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(57) Abstract

Methods for isolating K+Hnov genes are provided. The K+Hnov nucleic acid compositions find use in identifying homologous or related proteins and the DNA sequences encoding such proteins; in producing compositions that modulate the expression or function of the protein; and in studying associated physiological pathways. In addition, modulation of the gene activity in vivo is used for prophylactic and therapeutic purposes, such as identification of cell type based on expression, and the like.

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HUMAN POTASSIUM CHANNEL GENES

INTRODUCTION

Background

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lon channels are multi-subunit, membrane bound proteins critical for maintenance of cellular homeostasis in nearly all cell types. Channels are involved in a myriad of processes including modulation of action potentials, regulation of cardiac myocyte excitability, heart rate, vascular tone, neuronal signaling, activation and proliferation of T-cells, and insulin secretion from pancreatic islet cells. In humans, ion channels comprise extended gene families with hundreds, or perhaps thousands, of both closely related and highly divergent family members. The majority of known channels regulate the passage of sodium (Na*), chloride (Cl*), calcium (Ca**) and potassium (K*) ions across the cellular membrane.

Given their importance in maintaining normal cellular physiology, it is not surprising that ion channels have been shown to play a role in heritable human disease. Indeed, ion channel defects are involved in predisposition to epilepsy, cardiac arrhythmia (long QT syndrome), hypertension (Bartter's syndrome), cystic fibrosis, (defects in the CFTR chloride channel), several skeletal muscle disorders (hyperkalemic periodic paralysis, paramyotonia congenita, episodic ataxia) and congenital neural deafness (Jervell-Lange-Nielson syndrome), amongst others.

The potassium channel gene family is believed to be the largest and most diverse ion channel family. K* channels have critical roles in multiple cell types andpathways, and are the focus of significant investigation. Four human conditions, episodic ataxia with myokymia, long QT syndrome, epilepsy and Bartter's syndrome have been shown to be caused by defective K* ion channels. As the K* channel family is very diverse, and given that these proteins are critical components of virtually all cells, it is likely that abnormal K* channels will be involved in the etiology of additional renal, cardiovascular and central nervous system disorders of interest to the medical and pharmaceutical community.

The K* channel superfamily can be broadly classified into groups, based upon the number of transmembrane domain (TMD) segments in the mature

protein. The minK (IsK) gene contains a single TMD, and although not a channel by itself, minK associates with different K* channel subunits, such as KvLQT1 and HERG to modify the activity of these channels. The inward rectifying K+ channels (GIRK, IRK, CIR, ROMK) contain 2 TMD domains and a highly conserved pore domain. Twik-1 is a member of the newly emerging 4TMD K* channel subset. Twik-like channels (leak channels) are involved in maintaining the steady-state K* potentials across membranes and therefore the resting potential of the cell near the equilibrium potential for potassium (Duprat et al. (1997) EMBO J 16(17):5464-5471). These proteins are particularly intriguing targets for therapeutic regulation. The 6TMD, or Shaker-like channels, presently comprise the largest subset of known K* channels. The slopoke (slo) related channels, or Ca** regulated channels apparently have either 10 TMD, or 6 TMD with 4 additional hydrophobic domains.

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Four transmembrane domain, tandem pore domain K+ channels (4T/2P channels) represent a new family of potassium selective ion channels involved in the control of background membrane conductances. In mammals, five channels fitting the 4T/2P architecture have been described: TWIK, TREK, TASK-1, TASK-2 and TRAAK. The 4T/2P channels all have distinct characteristics, but are all thought to be involved in maintaining the steady-state K* potentials across membranes and therefore the resting potential of the cell near the equilibrium potential for potassium (Duprat et al. (1997) EMBO J 16(17):5464-5471). These proteins are particularly intriguing targets for therapeutic regulation. Within this group, TWIK-1, TREK-1 and TASK-1 and TASK-2 are widely distributed in many different tissues, while TRAAK is present exclusively in brain, spinal cord and retina. The 4T/2P channels have different physiologic properties; TREK-1 channels, are outwardly rectifying (Fink et al. (1996) EMBO J 15(24):6854-62), while TWIK-1 channels, are inwardly rectifying (Lesage et al. (1996) EMBO J 15(5):1004-11. TASK channels are regulated by changes in PH while TRAAK channels are stimulated by arachidonic acid (Reyes et al. (1998) JBC 273(47):30863-30869).

The degree of sequence homology between different K* channel genes is substantial. At the amino acid level, there is about 40% similarity between

different human genes, with distinct regions having higher homology, specifically the pore domain. It has been estimated that the K+ channel gene family contains approximately 10²-10³ individual genes. Despite the large number of potential genes, an analysis of public sequence databases and the scientific literature demonstrates that only a small number, approximately 20-30, have been identified. This analysis suggests that many of these important genes remain to be identified.

Potassium channels are involved in multiple different processes and are important regulators of homeostasis in nearly all cell types. Their relevance to basic cellular physiology and role in many human diseases suggests that pharmacological agents could be designed to specific channel subtypes and these compounds then applied to a large market (Bulman, D.E. (1997) Hum Mol Genet 6:1679-1685; Ackerman, M.J. and Clapham D.E. (1997) NEJM 336:1575-1586, Curran, M.E. (1998) Current Opinion in Biotechnology 9:565-572). The variety of therapeutic agents that modulate K+ channel activity reflects the diversity of physiological roles and importance of K+ channels in cellular function. A difficulty encountered in therapeutic use of therapeutic agents that modify K+ channel activity is that the presently available compounds tend to be non-specific and elicit both positive and negative responses, thereby reducing clinical efficacy. To facilitate development of specific compounds it is desirable to have further characterize novel K+ channels for use in *in vitro* and *in vivo* assays.

Relevant Literature

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A large body of literature exists in the general area of potassium channels. A review of the literature may be found in the series of books, "The Ion Channel Factsbook", volumes 1-4, by Edward C. Conley and William J. Brammar, Academic Press. An overview is provided of: extracellular ligand-gated ion channels (ISBN: 0121844501), intracellular ligand-gated channels (ISBN: 012184451X), Inward rectifier and intercellular channels (ISBN: 0121844528), and voltage gated channels (ISBN: 0121844536). Hille, B. (1992) "Ionic Channels of Excitable Membranes", 2nd Ed. Sunderland MA:Sinauer Associates, also reviews potassium channels.

Jan and Jan (1997) Annu. Rev. Neurosci. 20:91-123 review cloned potassium channels from eukaryotes and prokaryotes. Ackerman and Clapham (1997) N. Engl. J. Med. 336:1575-1586 discuss the basic science of ion channels in connection with clinical disease. Bulman (1997) Hum. Mol. Genet. 6:1679-1685 describe some phenotypic variation in ion channel disorders.

Stephan et al. (1994) Neurology 44:1915-1920 describe a pedigree segregating a myotonia with muscular hypertrophy and hyperirritability as an autosomal dominant trait (rippling muscle disease, Ricker et al. (1989) Arch. Neurol. 46405-408). Electromyography demonstrated that mechanical stimulation provoked electrically silent contractions. The responsible gene was localized to the distal end of the long arm of chromosome 1, in a 12-cM region near D1S235.

Type II pseudohypoaldosteronism is the designation used for a syndrome of chronic mineralocorticoid-resistant hyperkalemia with hypertension. The primary abnormality in type II PHA is thought to be a specific defect of the renal secretory mechanism for potassium, which limits the kaliuretic response to, but not the sodium and chloride reabsorptive effect of, mineralocorticoid. By analysis of linkage in families with autosomal dominant transmission, Mansfield *et al.* (1997) Nature Genet. 16:202-205 demonstrated locus heterogeneity of the trait, with linkage of the PHA2 gene to 1q31-q42 and 17p11-q21.

Sequences of four transmembrane, two pore potassium channels have been previously described. Reyes et al. (1998) J Biol Chem 273(47):30863-30869 discloses a pH sensitive channel. As with the related TASK-1 and TRAAK channels, the outward rectification is lost at high external K+ concentration. The TRAAK channel is described by Fink et al. (1998) EMBO J 17(12):3297-308. A cardiac two-pore channel is described in Kim et al. (1998) Circ Res 82(4):513-8. An open rectifier potassium channel with two pore domains in tandem and having a postsynaptic density protein binding sequence at the C terminal was cloned by Leonoudakis et al. (1998) J Neurosci 18(3):868-77.

The electrophysiological properties of Task channels are of interest, (Duprat et al. (1997) EMBO J 16:5464-71). TASK currents are K+-selective, instantaneous and non-inactivating. They show an outward rectification when external [K+] is low, which is not observed for high [K+]out, suggesting a lack of

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intrinsic voltage sensitivity. The absence of activation and inactivation kinetics as well as voltage independence are characteristic of conductances referred to as leak or background conductances. TASK is very sensitive to variations of extracellular pH in a narrow physiological range, a property probably essential for its physiological function, and suggests that small pH variations may serve a communication role in the nervous system.

SUMMARY OF THE INVENTION

Isolated nucleotide compositions and sequences are provided for K+Hnov genes. The K+Hnov nucleic acid compositions find use in identifying homologous or related genes; in producing compositions that modulate the expression or function of its encoded proteins; for gene therapy; mapping functional regions of the proteins; and in studying associated physiological pathways. In addition, modulation of the gene activity *in vivo* is used for prophylactic and therapeutic purposes, such as treatment of potassium channel defects, identification of cell type based on expression, and the like.

DESCRIPTION OF THE SPECIFIC EMBODIMENTS

Nucleic acid compositions encoding *K+Hnov* polypeptides are provided. They are used in identifying homologous or related genes; in producing compositions that modulate the expression or function of the encoded proteins; for gene therapy; mapping functional regions of the proteins; and in studying associated physiological pathways. The *K+Hnov* gene products are members of the potassium channel gene family, and have high degrees of homology to known potassium channels. The encoded polypeptides may be alpha subunits, which form the functional channel, or accessory subunits that act to modulate the channel activity.

CHARACTERIZATION OF K+HNOV

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The sequence data predict that the provided K+Hnov genes encode potassium channels. Table 1 summarizes the DNA sequences, corresponding SEQ ID NOs, chromosomal locations, and polymorphisms. The provided

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sequences may encode a predicted K*channel, e.g. voltage gated, inward rectifier, etc.; or a modulatory subunit.

Electrophysiologic characterization of ion channels is an important part of understanding channel function. Full length ion channel cDNAs may be combined with proper vectors to form expression constructs of each individual channel. Functional analyses of expressed channels can be performed in heterologous systems, or by expression in mammalian cell lines. For expression analyses in heterologous systems such as Xenopus oocytes, synthetic mRNA is made through in vitro transcription of each channel construct. mRNA is then injected, singly or in combination with interacting channel subunit mRNAs, into prepared oocytes and the cells allowed to express the channel for several days. Oocytes expressing the channel of interest are then analyzed by whole cell voltage clamp and patch clamp techniques.

To determine the properties of each channel when expressed in 15 mammalian cells expression vectors specific to this type of analyses may be constructed and the resultant construct used to transform the target cells (for example human embryonic kidney (HEK) cells). Both stable and transiently expressing lines may be studied using whole cell voltage clamp and patch clamp techniques. Data obtained from EP studies includes, but is not limited to: current profiles elicited by depolarization and hyperpolarization, current-voltage (I-V) relationships, voltage dependence of activation, biophysical kinetics of channel activation, deactivation, and inactivation, reversal potential, ion selectivity, gating properties and sensitivity to channel antagonists and agonists.

Heterologous or mammalian cell lines expressing the novel channels can be used to characterize small molecules and drugs which interact with the channel. The same experiments can be used to assay for novel compounds which interact with the expressed channels.

In many cases the functional ion channel formed by K+Hnov polypeptides will be heteromultimers. Heteromultimers are known to form between different voltage gated, outward rectifying potassium channel α subunits, generally comprising four subunits, and frequently associated with auxiliary, β subunits. Typically such α subunits share a six-transmembrane domain structure (S1-S6),

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with one highly positively charged domain (S4) and a pore region situated between S5 and S6. Examples of such subunits are K+Hnov4, K+Hnov9, and K+Hnov12. Channels are also formed by mutimerization of subunits of the two transmembrane and one pore architecture. It is predicted that two subunits of K+Hnov49 or K+Hnov59 will be required to form a functional channel.

Heteromultimers of greatest interest are those that form between subunits expressed in the same tissues, and are a combination of subunits from the same species. In addition, the formation of multimers between the subject polypeptides and subunits that form functional channels are of particular interest. The resulting channel may have decreased or increased conductance relative to a homomultimer, and may be altered in response to beta subunits or other modulatory molecules.

Known voltage gated K+ channel α subunits include Kv1.1-1.8 (Gutman et al. (1993) Sem. Neurosci. 5:101-106); Kv2.1-2.2; Kv3.1-3.4; Kv4.1-4.3; Kv5.1; Kv6.1; Kv7.1; Kv8.1; Kv9.1-9.2. The subunits capable of forming ion inducing channels include all of those in the Kv1 through Kv4; and Kv7 families. The Kv5.1, Kv6.1, Kv8.1 and Kv9.1-9.2 subunits may be electrically silent, but functional in modifying the properties in heteromultimers.

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TABLE 1

	ard rectifying	channel						K+ channel	hannel	hannel				3		
ATP-sensitive inward rectifying		Voltage gated K+ channel				:		Delayed rectifying K+ channel	Voltage gated K+ channel	Voltage gated K+ channel		. :	modulatory subunit			modulatory subunit
	2q37	unknown			:			2p23	8q23	Xp21			13q14	3.4		18q12
	Alternative poly(A) tail: 1236, 2395	A312C	T335C	A377G	T344C	A401G	CA410-411GG (Ala/Thr)		Alternative poly(A) tail: 2304	C321T (Pro/Leu)	A375G (Glu/Gly)	C407T (Leu/Phe)	Alternative poly(A) tail: 1427	A689G (Gly/Arg)		1365A (IIe/Asn)
	SEQ ID NO:2	SEQ ID NO:4						SEQ ID NO:6	SEQ ID NO:8	SEQ ID NO:10			SEQ ID NO:12		11 OIL OIL OIL	SEC ID NO. 14
	SEQ ID NO:1	SEQ ID NO:3			: : : : : : : : : : : : : : : : : : : :	· · · · · · · · · · · · · · · · · · ·		SEQ ID NO:5	SEQ ID NO:7	SEQ ID NO:9	-		SEQ ID NO:11		SEO ID NO:13	SEG 10 140. 13
	K+Hnov1	K+Hnov4						K+Hnov6	K+Hnov9	K+Mnov12			K+Hnov15		K+Hnov27	17.1.111012.1

OMS וב כבי פעד

K+Hnov 11	SEQ ID NO:17	SEQ ID NO:18	N/A	N/A	Human ortholog of murine gene, 6
			,		transmembrane dominas, voltage
					gated, delayed rectifier K+ channel
K+Hnov 14	SEQ ID NO:19	SEQ ID NO:20	C3168T	12q14	6 transmembrane domain, voltage
K+Hnov28	SEQ ID NO:21-24	SEQ ID NO.25	4 alternative 5' splices	3q29	Modulatory subunit
K+Hnov42	SEQ ID NO:26	SEQ ID NO:27	G1162A, T1460A, T2496A	8q11	Homology to K+ channel protein of C. elegans
K+Hnov44	SEQ ID NO:28-29	SEQ ID NO.30	NA	22p13	beta-subunit.
K'Hnov49	SEQ ID NO:80	SEG ID NO:81	(ATCT), repeats in the 3' UTR sequence, starting at position 2166	1941	4T2P channel, linked to the disease loci for rippling muscle disease 1 (RMD1), and type II pseudohypopaldosteronism
K'Hnov59	SEQ ID NO:82	SEQ ID NO:83	WA .	chr19	4T2P channel

K+HNOV NUCLEIC ACID COMPOSITIONS

As used herein, the term "K+Hnov" is generically used to refer to any one of the provided genetic sequences listed in Table 1. Where a specific K+Hnov sequence is intended, the numerical designation, e.g. K49 or K59, will be added. Nucleic acids encoding K+Hnov potassium channels may be cDNA or genomic DNA or a fragment thereof. The term "K+Hnov gene" shall be intended to mean the open reading frame encoding any of the provided K+Hnov polypeptides, introns, as well as adjacent 5' and 3' non-coding nucleotide sequences involved in the regulation of expression, up to about 20 kb beyond the coding region, but possibly further in either direction. The gene may be introduced into an appropriate vector for extrachromosomal maintenance or for integration into a host genome.

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The term "cDNA" as used herein is intended to include all nucleic acids that share the arrangement of sequence elements found in native mature mRNA species, where sequence elements are exons and 3' and 5' non-coding regions. Normally mRNA species have contiguous exons, with the intervening introns, when present, removed by nuclear RNA splicing, to create a continuous open reading frame encoding a K+Hnov protein.

A genomic sequence of interest comprises the nucleic acid present between the initiation codon and the stop codon, as defined in the listed sequences, including all of the introns that are normally present in a native chromosome. It may further include the 3' and 5' untranslated regions found in the mature mRNA. It may further include specific transcriptional and translational regulatory sequences, such as promoters, enhancers, etc., including about 1 kb, but possibly more, of flanking genomic DNA at either the 5' or 3' end of the transcribed region. The genomic DNA may be isolated as a fragment of 100 kbp or smaller; and substantially free of flanking chromosomal sequence. The genomic DNA flanking the coding region, either 3' or 5', or internal regulatory sequences as sometimes found in introns, contains sequences required for proper tissue and stage specific expression.

The sequence of the 5' flanking region may be utilized for promoter elements, including enhancer binding sites, that provide for developmental regulation in tissues where *K+Hnov* genes are expressed. The tissue specific expression is useful for determining the pattern of expression, and for providing promoters that mimic the native pattern of expression. Naturally occurring polymorphisms in the promoter regions are useful for determining natural variations in expression, particularly those that may be associated with disease.

Alternatively, mutations may be introduced into the promoter regions to determine the effect of altering expression in experimentally defined systems. Methods for the identification of specific DNA motifs involved in the binding of transcriptional factors are known in the art, e.g. sequence similarity to known binding motifs, gel retardation studies, etc. For examples, see Blackwell et al. (1995) Mol Med 1: 194-205; Mortlock et al. (1996) Genome Res. 5: 327-33; and Joulin and Richard-Foy (1995) Eur J Biochem 232: 620-626.

The regulatory sequences may be used to identify *cis* acting sequences required for transcriptional or translational regulation of *K+Hnov* expression, especially in different tissues or stages of development, and to identify *cis* acting sequences and *trans* acting factors that regulate or mediate *K+Hnov* expression. Such transcription or translational control regions may be operably linked to a *K+Hnov* gene in order to promote expression of wild type or altered *K+Hnov* or other profeins of interest in cultured cells, or in embryonic, fetal or adult tissues, and for gene therapy.

The nucleic acid compositions of the subject invention may encode all or a part of the subject polypeptides. Double or single stranded fragments may be obtained of the DNA sequence by chemically synthesizing oligonucleotides in accordance with conventional methods, by restriction enzyme digestion, by PCR amplification, etc. For the most part, DNA fragments will be of at least 15 nt, usually at least 18 nt or 25 nt, and may be at least about 50 nt. Such small DNA fragments are useful as primers for PCR, hybridization screening probes, etc. Larger DNA fragments, i.e. greater than 100 nt are useful for production of the encoded polypeptide. For use in amplification reactions, such as PCR, a pair of

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primers will be used. The exact composition of the primer sequences is not critical to the invention, but for most applications the primers will hybridize to the subject sequence under stringent conditions, as known in the art. It is preferable to choose a pair of primers that will generate an amplification product of at least about 50 nt, preferably at least about 100 nt. Algorithms for the selection of primer sequences are generally known, and are available in commercial software packages. Amplification primers hybridize to complementary strands of DNA, and will prime towards each other.

The K+Hnov genes are isolated and obtained in substantial purity, generally as other than an intact chromosome. Usually, the DNA will be obtained substantially free of other nucleic acid sequences that do not include a K+Hnov sequence or fragment thereof, generally being at least about 50%, usually at least about 90% pure and are typically "recombinant", i.e. flanked by one or more nucleotides with which it is not normally associated on a naturally occurring chromosome.

The DNA may also be used to identify expression of the gene in a biological specimen. The manner in which one probes cells for the presence of particular nucleotide sequences, as genomic DNA or RNA, is well established in the literature and does not require elaboration here. DNA or mRNA is isolated from a cell sample. The mRNA may be amplified by RT-PCR, using reverse transcriptase to form a complementary DNA strand, followed by polymerase chain reaction amplification using primers specific for the subject DNA sequences. Alternatively, the mRNA sample is separated by gel electrophoresis, transferred to a suitable support, e.g. nitrocellulose, nylon, etc., and then probed with a fragment of the subject DNA as a probe. Other techniques, such as oligonucleotide ligation assays, in situ hybridizations, and hybridization to DNA probes arrayed on a solid chip may also find use. Detection of mRNA hybridizing to the subject sequence is indicative of K+Hnov gene expression in the sample.

The sequence of a K+Hnov gene, including flanking promoter regions and cooling regions, may be mutated in various ways known in the art to generate targeted changes in promoter strength, sequence of the encoded protein, etc.

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The DNA sequence or protein product of such a mutation will usually be substantially similar to the sequences provided herein, i.e. will differ by at least one nucleotide or amino acid, respectively, and may differ by at least two but not more than about ten nucleotides or amino acids. The sequence changes may be substitutions, insertions or deletions. Deletions may further include larger changes, such as deletions of a domain or exon. Other modifications of interest include epitope tagging, e.g. with the FLAG system, HA, etc. For studies of subcellular localization, fusion proteins with green fluorescent proteins (GFP) may be used.

Techniques for *in vitro* mutagenesis of cloned genes are known. Examples of protocols for site specific mutagenesis may be found in Gustin *et al.*, *Biotechniques* 14:22 (1993); Barany, *Gene* 37:111-23 (1985); Colicelli *et al.*, *Mol Gen Genet* 199:537-9 (1985); and Prentki *et al.*, *Gene* 29:303-13 (1984). Methods for site specific mutagenesis can be found in Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, CSH Press 1989, pp. 15.3-15.108; Weiner *et al.*, *Gene* 126:35-41 (1993); Sayers *et al.*, *Biotechniques* 13:592-6 (1992); Jones and Winistorfer, *Biotechniques* 12:528-30 (1992); Barton *et al.*, *Nucleic Acids Res* 18:7349-55 (1990); Marotti and Tomich, *Gene Anal Tech* 6:67-70 (1989); and Zhu, *Anal Biochem* 177:120-4 (1989). Such mutated genes may be used to study structure-function relationships of *K+Hnov*, or to alter properties of the protein that affect its function or regulation.

Homologs and orthologs of K+Hnov genes are identified by any of a number of methods. A fragment of the provided cDNA may be used as a hybridization probe against a cDNA library from the target organism of interest, where low stringency conditions are used. The probe may be a large fragment, or one or more short degenerate primers. Nucleic acids having sequence similarity are detected by hybridization under low stringency conditions, for example, at 50°C and 6XSSC (0.9 M sodium chloride/0.09 M sodium citrate) and remain bound when subjected to washing at 55°C in 1XSSC (0.15 M sodium chloride/0.015 M sodium citrate). Sequence identity may be determined by hybridization under stringent conditions, for example, at 50°C or higher and

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0.1XSSC (15 mM sodium chloride/01.5 mM sodium citrate). Nucleic acids having a region of substantial identity to the provided K+Hnov sequences, e.g. allelic variants, genetically altered versions of the gene, etc., bind to the provided K+Hnov sequences under stringent hybridization conditions. By using probes, particularly labeled probes of DNA sequences, one can isolate homologous or related genes. The source of homologous genes may be any species, e.g. primate species, particularly human; rodents, such as rats and mice, canines, felines, bovines, ovinas, equines, yeast, nematodes, etc.

Between mammalian species, e.g. human and mouse, homologs have 10 substantial sequence similarity, i.e. at least 75% sequence identity between nucleotide sequences, in some cases 80 or 90% sequence identity, and may be as high as 95% sequence identity between closely related species. Sequence similarity is calculated based on a reference sequence, which may be a subset of a larger sequence, such as a conserved motif, coding region, flanking region, etc. 15 A reference sequence will usually be at least about 18 nt long, more usually at least about 30 nt long, and may extend to the complete sequence that is being compared. Algorithms for sequence analysis are known in the art, such as BLAST, described in Altschul et al. (1990), J. Mol. Biol. 215:403-10. In general, variants of the invention have a sequence identity greater than at least about 65%, preferably at least about 75%, more preferably at least about 85%, and may be greater than at least about 90% or more as determined by the Smith-Waterman homology search algorithm as implemented in MPSRCH program (Oxford Molecular). Exemplary search parameters for use with the MPSRCH program in order to identify sequences of a desired sequence identity are as follows: gap open penalty: 12; and gap extension penalty: 1. 25

K+HNOV POLYPEPTIDES

The subject nucleic acid sequences may be employed for producing all or portions of K+Hnov polypeptides. For expression, an expression cassette may be employed. The expression vector will provide a transcriptional and translational initiation region, which may be inducible or constitutive, where the coding region

is operably linked under the transcriptional control of the transcriptional initiation region, and a transcriptional and translational termination region. These control regions may be native to a K+Hnov gene; or may be derived from exogenous sources.

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The peptide may be expressed in prokaryotes or eukaryotes in accordance with conventional ways, depending upon the purpose for expression. For large scale production of the protein, a unicellular organism, such as *E. coli, B. subtilis, S. cerevisiae*, insect cells in combination with baculovirus vectors, or cells of a higher organism such as vertebrates, particularly mammals, e.g. COS 7 cells, may be used as the expression host cells. In some situations, it is desirable to express the *K+Hnov* gene in eukaryotic cells, where the *K+Hnov* protein will benefit from native folding and post-translational modifications. Small peptides can also be synthesized in the laboratory. Peptides that are subsets of the complete *K+Hnov* sequence may be used to identify and investigate parts of the protein important for function, or to raise antibodies directed against these regions.

Fragments of interest include the transmembrane and pore domains, the signal sequences, regions of interaction between subunits, etc. Such domains will usually include at least about 20 amino acids of the provided sequence, more usually at least about 50 amino acids, and may include 100 amino acids or more, up to the complete domain. Binding contacts may be comprised of non-contiguous sequences, which are brought into proximity by the tertiary structure of the protein. The sequence of such fragments may be modified through manipulation of the coding sequence, as described above. Truncations may be performed at the carboxy or amino terminus of the fragment, e.g. to determine the minimum sequence required for biological activity.

With the availability of the protein or fragments thereof in large amounts, by employing an expression host, the protein may be isolated and purified in accordance with conventional ways. A lysate may be prepared of the expression host and the lysate purified using HPLC, exclusion chromatography, gel electrophoresis, affinity chromatography, or other purification technique. The

purified protein will generally be at least about 80% pure, preferably at least about 90% pure, and may be up to and including 100% pure. Pure is intended to mean free of other proteins, as well as cellular debris.

The expressed K+Hnov polypeptides are useful for the production of antibodies, where short fragments provide for antibodies specific for the particular polypeptide, and larger fragments or the entire protein allow for the production of antibodies over the surface of the polypeptide. Antibodies may be raised to the wild-type or variant forms of K+Hnov. Antibodies may be raised to isolated peptides corresponding to specific domains, e.g. the pore domain and the transmembrane domain, or to the native protein.

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Antibodies are prepared in accordance with conventional ways, where the expressed polypeptide or protein is used as an immunogen, by itself or conjugated to known immunogenic carriers, e.g. KLH, pre-S HBsAg, other viral or eukaryotic proteins, or the like. Various adjuvants may be employed, with a series of injections, as appropriate. For monoclonal antibodies, after one or more booster injections, the spleen is isolated, the lymphocytes immortalized by cell fusion, and then screened for high affinity antibody binding. The immortalized cells, i.e. hybridomas, producing the desired antibodies may then be expanded. For further description, see Monoclonal Antibodies: A Laboratory Manual, Harlow and Lane eds., Cold Spring Harbor Laboratories, Cold Spring Harbor, New York, 1988. If desired, the mRNA encoding the heavy and light chains may be isolated and mutagenized by cloning in E. coli, and the heavy and light chains mixed to further enhance the affinity of the antibody. Alternatives to in vivo immunization as a method of raising antibodies include binding to phage "display" libraries, usually in conjunction with in vitro affinity maturation.

K+HNOV GENOTYPING

The subject nucleic acid and/or polypeptide compositions may be used to genotyping and other analysis for the presence of polymorphisms in the sequence, or variation in the expression of the subject genes. Genotyping may be performed to determine whether a particular polymorphisms is associated with

a disease state or genetic predisposition to a disease state, particularly diseases associated with defects in excitatory properties of cells, e.g. cardiac, muscle, renal and neural cells. Disease of interest include rippling muscle disease, and type II psuedohypoaldosteronism.

Clinical disorders associated with K+ channel defects include long-QT syndrome; a congenital disorder affecting 1 in 10,000-15,000. Affected individuals have a prolonged QT interval in the electrocardiogram due to a delayed repolarization of the ventricle. Genetic linkage analyses identified two loci for long QT syndrome, LQT1, in 11p15.5 and LQT2, in 7q35-36. Positional cloning techniques identified the novel K+ channel KvLQT1 on chromosome 11 while candidate gene analysis identified causative mutations in the HERG K+ channel for LQT2.

The weaver mouse exhibits several abnormal neurological symptoms, including severe ataxia, loss of granule cell neurons in the cerebellum and dopaminergic cells in the substantia nigra, as well as seizures and male infertility. A G-protein-coupled K+ channel having a mutation in the conserved pore domain has been determined to cause the disease. The pancreatic-islet \(\mathcal{G}\)-cell ATP-sensitive K+ channel (KATP) is composed of two subunits, the sulfonylurea receptor (SUR) and the inward rectifier K+ channel Kir6.2. Mutations in both SUR and Kir6.2 have been identified in patients with persistent hyperinsulinemic hypoglycemia of infancy, which is caused by unregulated secretion of insulin.

Genotyping may also be performed for pharmacogenetic analysis to assess the association between an individual's genotype and that individual's ability to react to a therapeutic agent. Differences in target sensitivity can lead to toxicity or therapeutic failure. Relationships between polymorphisms in channel expression or specificity can be used to optimize therapeutic dose administration.

Genetic polymorphisms are identified in the K+Hnov gene (examples are listed in table 1), e.g. the repeat variation in the 3' UTR of K49. Nucleic acids comprising the polymorphic sequences are used to screen patients for altered reactivity and adverse side effects in response to drugs that act on K+ channels.

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K+Hnov genotyping is performed by DNA or RNA sequence and/or hybridization analysis of any convenient sample from a patient, e.g. biopsy material, blood sample, scrapings from cheek, etc. A nucleic acid sample from an individual is analyzed for the presence of polymorphisms in K+Hnov. particularly those that affect the activity, responsiveness or expression of K+Hnov. Specific sequences of interest include any polymorphism that leads to changes in basal expression in one or more tissues, to changes in the modulation of K+Hnov expression, or alterations in K+Hnov specificity and/or activity.

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The effect of a polymorphism in K+Hnov gene sequence on the response to a particular agent may be determined by in vitro or in vivo assays. Such assays may include monitoring during clinical trials, testing on genetically defined cell lines, etc. The response of an individual to the agent can then be predicted by determining the K+Hnov genotype with respect to the polymorphism. Where there is a differential distribution of a polymorphism by racial background, 15 guidelines for drug administration can be generally tailored to a particular ethnic group.

Biochemical studies may be performed to determine whether a sequence polymorphism in a K+Hnov coding region or control regions is associated with disease, for example the association of K+Hnov 9 with idiopathic generalized epilepsy. Disease associated polymorphisms may include deletion or truncation of the gene, mutations that after expression level, that affect the electrical activity of the channel, etc.

A number of methods are available for analyzing nucleic acids for the presence of a specific sequence. Where large amounts of DNA are available, genomic DNA is used directly. Alternatively, the region of interest is cloned into a suitable vector and grown in sufficient quantity for analysis. The nucleic acid may be amplified by conventional techniques, such as the polymerase chain reaction (PCR), to provide sufficient amounts for analysis. The use of the polymerase chain reaction is described in Saiki et al. (1985) Science 239:487, and a review of current techniques may be found in Sambrook et al. Molecular Cloning: A Laboratory Manual, CSH Press 1989, pp.14.2-14.33. Amplification may be used

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to determine whether a polymorphism is present, by using a primer that is specific for the polymorphism. Alternatively, various methods are known in the art that utilize oligonucleotide ligation as a means of detecting polymorphisms, for examples see Riley et al. (1990) N.A.R. 18:2887-2890; and Delahunty et al. (1996) Am. J. Hum. Genet.58:1239-1246.

A detectable label may be included in an amplification reaction. Suitable labels include fluorochromes, e.g. fluorescein isothiocyanate (FITC), rhodamine, Texas Red, phycoerythrin, allophycocyanin, 6-carboxyfluorescein (6-FAM), 2',7'-dimethoxy-4',5'- dichloro-6-carboxyfluorescein (JOE), 6-carboxy-X-rhodamine (ROX), 6-carboxy-2',4',7',4,7- hexachlorofluorescein (HEX), 5-carboxyfluorescein (5-FAM) or N,N,N',N'-tetramethyl-6- carboxyrhodamine (TAMRA), radioactive labels, e.g. 32P, 35S, 3H; etc., The label may be a two stage system, where the amplified DNA is conjugated to biotin, haptens, etc. having a high affinity binding partner, e.g. avidin, specific antibodies, etc., where the binding partner is conjugated to a detectable label. The label may be conjugated to one or both of the primers. Alternatively, the pool of nucleotides used in the amplification is labeled, so as to incorporate the label into the amplification product.

The sample nucleic acid, e.g. amplified or cloned fragment, is analyzed by one of a number of methods known in the art. The nucleic acid may be sequenced by dideoxy or other methods. Hybridization with the variant sequence may also be used to determine its presence, by Southern blots, dot blots, etc. The hybridization pattern of a control and variant sequence to an array of oligonucleotide probes immobilised on a solid support, as described in U.S. 5,445,934, or in WO95/35505, may also be used as a means of detecting the presence of variant sequences. Single strand conformational polymorphism (SSCP) analysis, denaturing gradient gel electrophoresis (DGGE), mismatch cleavage detection, and heteroduplex analysis in gel matrices are used to detect conformational changes created by DNA sequence variation as alterations in electrophoretic mobility. Alternatively, where a polymorphism creates or destroys a recognition site for a restriction endonuclease (restriction fragment length polymorphism, RFLP), the sample is digested with that endonuclease, and the

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products size fractionated to determine whether the fragment was digested. Fractionation is performed by gel or capillary electrophoresis, particularly acrylamide or agarose gels.

In one embodiment of the invention, an array of oligonucleotides are provided, where discrete positions on the array are complementary to one or more of the provided sequences, e.g. oligonucleotides of at least 12 nt, frequently 20 nt, or larger, and including the sequence flanking a polymorphic position in a K*Hnov sequence; coding sequences for different K*Hnov channels, panels of ion channels comprising one or more of the provided K* channels; etc. Such an array may comprise a series of oligonucleotides, each of which can specifically hybridize to a different polymorphism. For examples of arrays, see Hacia et al. (1996) Nature Genetics 14:441-447; Lockhart et al. (1996) Nature Biotechnol. 14:1675-1680; and De Risi et al. (1996) Nature Genetics 14:457-460.

Screening for polymorphisms in K+Hnov may be based on the functional or antigenic characteristics of the protein. Protein truncation assays are useful in detecting deletions that may affect the biological activity of the protein. Various immunoassays designed to detect polymorphisms in K+Hnov proteins may be used in screening. Where many diverse genetic mutations lead to a particular disease phenotype, functional protein assays have proven to be effective screening tools. The activity of the encoded K+Hnov protein as a potassium channel may be determined by comparison with the wild-type protein.

Antibodies specific for a K+Hnov may be used in staining or in immunoassays. Samples, as used herein, include biological fluids such as semen, blood, cerebrospinal fluid, tears, saliva, lymph, dialysis fluid and the like; organ or tissue culture derived fluids; and fluids extracted from physiological tissues. Also included in the term are derivatives and fractions of such fluids. The cells may be dissociated, in the case of solid tissues, or tissue sections may be analyzed. Alternatively a lysate of the cells may be prepared.

Diagnosis may be performed by a number of methods to determine the absence or presence or altered amounts of normal or abnormal K+Hnov polypeptides in patient cells. For example, detection may utilize staining of cells

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or histological sections, performed in accordance with conventional methods. The antibodies of interest are added to the cell sample, and incubated for a period of time sufficient to allow binding to the epitope, usually at least about 10 minutes. The antibody may be labeled with radioisotopes, enzymes, fluorescers, chemiluminescers, or other labels for direct detection. Alternatively, a second stage antibody or reagent is used to amplify the signal. Such reagents are well known in the art. For example, the primary antibody may be conjugated to biotin, with horseradish peroxidase-conjugated avidin added as a second stage reagent. Alternatively, the secondary antibody conjugated to a flourescent compound, e.g. flourescein, rhodamine, Texas red, etc. Final detection uses a substrate that undergoes a color change in the presence of the peroxidase. The absence or presence of antibody binding may be determined by various methods, including flow cytometry of dissociated cells, microscopy, radiography, scintillation counting, etc.

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MODULATION OF GENE EXPRESSION

The K+Hnov genes, gene fragments, or the encoded protein or protein fragments are useful in gene therapy to treat disorders associated with K+Hnov defects. Expression vectors may be used to introduce the K+Hnov gene into a cell. Such vectors generally have convenient restriction sites located near the promoter sequence to provide for the insertion of nucleic acid sequences. Transcription cassettes may be prepared comprising a transcription initiation region, the target gene or fragment thereof, and a transcriptional termination region. The transcription cassettes may be introduced into a variety of vectors, e.g. plasmid; retrovirus, e.g. lentivirus; adenovirus; and the like, where the vectors are able to transiently or stably be maintained in the cells, usually for a period of at least about several days to several weeks.

The gene or K+Hnov protein may be introduced into tissues or host cells by any number of routes, including viral infection, microinjection, or fusion of vesicles. Jet injection may also be used for intramuscular administration, as

described by Furth et al. (1992) Anal Biochem 205:365-368. The DNA may be coated onto gold microparticles, and delivered intradermally by a particle bombardment device, or "gene gun" as described in the literature (see, for example, Tang et al. (1992) Nature 356:152-154), where gold microprojectiles are coated with the K+Hnov or DNA, then bombarded into skin ceils.

Antisense molecules can be used to down-regulate expression of K+Hnov in cells. The anti-sense reagent may be antisense oligonuclectides (ODN), particularly synthetic ODN having chemical modifications from native nucleic acids, or nucleic acid constructs that express such anti-sense molecules as RNA. The antisense sequence is complementary to the mRNA of the targeted gene, and inhibits expression of the targeted gene products. Antisense molecules inhibit gene expression through various mechanisms, e.g. by reducing the amount of mRNA available for translation, through activation of RNAse H, or steric hindrance. One or a combination of antisense molecules may be administered, where a combination may comprise multiple different sequences.

Antisense molecules may be produced by expression of all or a part of the target gene sequence in an appropriate vector, where the transcriptional initiation is oriented such that an antisense strand is produced as an RNA molecule. Alternatively, the antisense molecule is a synthetic oligonucleotide. Antisense oligonucleotides will generally be at least about 7, usually at least about 12, more usually at least about 20 nucleotides in length, and not more than about 500, usually not more than about 50, more usually not more than about 35 nucleotides in length, where the length is governed by efficiency of inhibition, specificity, including absence of cross-reactivity, and the like. It has been found that short oligonucleotides, of from 7 to 8 bases in length, can be strong and selective inhibitors of gene expression (see Wagner et al. (1996) Nature Biotechnology 14:840-844).

A specific region or regions of the endogenous sense strand mRNA sequence is chosen to be complemented by the antisense sequence. Selection of a specific sequence for the oligonucleotide may use an empirical method, where several candidate sequences are assayed for inhibition of expression of

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the target gene in an in vitro or animal model. A combination of sequences may also be used, where several regions of the rnRNA sequence are selected for antisense complementation.

Antisense oligonucleotides may be chemically synthesized by methods known in the art (see Wagner et al. (1993) supra. and Milligan et al., supra.) Preferred oligonucleotides are chemically modified from the native phosphodiester structure, in order to increase their intracellular stability and binding affinity. A number of such modifications have been described in the literature, which alter the chemistry of the backbone, sugars or heterocyclic 10 bases.

Among useful changes in the backbone chemistry are phosphorothioates; phosphorodithioates, where both of the non-bridging oxygens are substituted with sulfur; phosphoroamidites; alkyl phosphotriesters and boranophosphates. Achiral phosphate derivatives include 3'-O'-5'-S-phosphorothioate, 3'-S-5'-Ophosphorothicate, 3'-CH2-5'-O-phosphonate and 3'-NH-5'-O-phosphoroamidate. Peptide nucleic acids replace the entire ribose phosphodiester backbone with a peptide linkage. Sugar modifications are also used to enhance stability and -affinity. The α -anomer of deoxyribose may be used, where the base is inverted with respect to the natural β-anomer. The 2'-OH of the ribose sugar may be altered to form 2'-O-methyl or 2'-O-allyl sugars, which provides resistance to degradation without comprising affinity. Modification of the heterocyclic bases must maintain proper base pairing. Some useful substitutions include deoxyuridine for deoxythymidine; 5-methyl-2'-deoxycytidine and 5-bromo-2'deoxycytidine for deoxycytidine. 5- propynyl-2'-deoxyuridine and 5-propynyl-2'-25 deoxycytidine have been shown to increase affinity and biological activity when substituted for deoxythymidine and deoxycytidine, respectively.

As an alternative to anti-sense inhibitors, catalytic nucleic acid compounds, e.g. ribozymes, anti-sense conjugates, etc. may be used to inhibit gene expression. Ribozymes may be synthesized in vitro and administered to the 20 patient, or may be encoded on an expression vector, from which the ribozyme is synthesized in the targeted cell (for example, see International patent application

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WO 9523225, and Beigelman et al. (1995) Nucl. Acids Res 23:4434-42). Examples of oligonucleotides with catalytic activity are described in WO 9506764. Conjugates of anti-sense ODN with a metal complex, e.g. terpyridylCu(II), capable of mediating mRNA hydrolysis are described in Bashkin et al. (1995) Appl 5 Biochem Biotechnol 54:43-56.

GENETICALLY ALTERED CELL OR ANIMAL MODELS FOR K+HNOV FUNCTION

The subject nucleic acids can be used to generate transgenic animals or site specific gene modifications in cell lines. Transgenic animals may be made through homologous recombination, where the normal K+Hnov locus is altered. Alternatively, a nucleic acid construct is randomly integrated into the genome. Vectors for stable integration include plasmids, retroviruses and other animal viruses, YACs, and the like.

The modified cells or animals are useful in the study of K+Hnov function and regulation. For example, a series of small deletions and/or substitutions may be made in the K+Hnov gene to determine the role of different transmembrane domains in forming multimeric structures, ion channels, etc. Of interest are the use of K+Hnov to construct transgenic animal models for epilepsy and other neurological defects, where expression of K+Hnov is specifically reduced or absent. Specific constructs of interest include anti-sense K+Hnov, which will block K+Hnov expression, expression of dominant negative K+Hnov mutations, etc. One may also provide for expression of the K+Hnov gene or variants thereof in cells or tissues where it is not normally expressed or at abnormal times of development.

DNA constructs for homologous recombination will comprise at least a portion of the K+Hnov gene with the desired genetic modification, and will include regions of homology to the target locus. DNA constructs for random integration need not include regions of homology to mediate recombination. Conveniently, markers for positive and negative selection are included. Methods for generating cells having targeted gene modifications through homologous recombination are

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known in the art. For various techniques for transfecting mammalian cells, see Keown et al. (1990) Methods in Enzymology 185:527-537.

For embryonic stem (ES) cells, an ES cell line may be employed, or embryonic cells may be obtained freshly from a host, e.g. mouse, rat, guinea pig, etc. Such cells are grown on an appropriate fibroblast-feeder layer or grown in the presence of leukemia inhibiting factor (LIF). When ES or embryonic cells have been transformed, they may be used to produce transgenic animals. After transformation, the cells are plated onto a feeder layer in an appropriate medium. Cells containing the construct may be detected by employing a selective medium. After sufficient time for colonies to grow, they are picked and analyzed for the occurrence of homologous recombination or integration of the construct. Those colonies that are positive may then be used for embryo manipulation and blastocyst injection. Blastocysts are obtained from 4 to 6 week old superovulated females. The ES cells are trypsinized, and the modified cells are injected into the blastocoel of the blastocyst. After injection, the blastocysts are returned to each uterine horn of pseudopregnant females. Females are then allowed to go to term and the resulting offspring screened for the construct. By providing for a different phenotype of the blastocyst and the genetically modified cells, chimeric progeny can be readily detected.

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The chimeric animals are screened for the presence of the modified gene and males and females having the modification are mated to produce homozygous progeny. If the gene alterations cause lethality at some point in development, tissues or organs can be maintained as allogeneic or congenic grafts or transplants, or in *in vitro* culture. The transgenic animals may be any non-human mammal, such as laboratory animals, domestic animals, etc. The transgenic animals may be used in functional studies, drug screening, etc., e.g. to determine the effect of a candidate drug on Ras or related gene activation, oncogenesis, etc.

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TESTING OF K+HNOV FUNCTION and RESPONSES

Potassium channels such as K+Hnov polypeptides are involved in multiple biologically important processes. Pharmacological agents designed to affect only specific channel subtypes are of particular interest. Presently available compounds tend to be non-specific and elicit both positive and negative responses, thereby reducing clinical efficacy.

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The subject polypeptides may be used in *in vitro* and *in vivo* models to test the specificity of novel compounds, and of analogs and derivatives of compounds known to act on potassium channels. Numerous pharmacological agents have profound affects on K+ channel activity. As examples, Sotalol (BETAPACE) is a class III antiarrhythmic drug that prolongs cardiac action potentials by inhibiting delayed rectifier K+ channels. Sulfonylurea drugs, such as Glipizide (GLUCOTROL) and Tolazamide (TOLAMIDE) function as antidiabetic drugs by blocking ATP-sensitive K+ channels present in pancreatic islet cells, thereby regulating insulin secretion. Diazoxide (HYPERSTAT IV) is an antihypertensive drug that activates ATP-sensitive K+ channels, resulting in the relaxation of vascular smooth muscle. There are several other examples of drugs that have antidiabetic, antihypertensive, or antiarrhythmic activities. A number of drugs that activate K+ channels that have been proposed as coronary vasodilators for the treatment of both vasospastic and chronic stable angina.

The availability of multiple K+ channel subunits allows *in vitro* reconstruction of functional channels, which may comprise different alpha and beta subunits. The individual components may be modified by sequence deletion, substitution, *etc.* to determine the functional role of specific domains.

Drug screening may be performed using an *in vitro* model, a genetically altered cell or animal, or purified K+Hnov protein, either as monomers, homomultimers or hetermultimers. One can identify ligands or substrates that bind to, modulate or mimic the action of K+Hnov. Drug screening identifies agents that provide a replacement for K+Hnov function in abnormal cells. Of particular interest are screening assays for agents that have a low toxicity for human cells. A wide variety of assays may be used for this purpose, including

monitoring cellular excitation and conductance, labeled *in vitro* protein-protein binding assays, electrophoretic mobility shift assays, immunoassays for protein binding, and the like. The purified protein may also be used for determination of three-dimensional crystal structure, which can be used for modeling intermolecular interactions.

The term "agent" as used herein describes any molecule, e.g. protein or pharmaceutical, with the capability of altering or mimicking the physiological function of K+Hnov polypeptide. Generally a plurality of assay mixtures are run in parallel with different agent concentrations to obtain a differential response to the various concentrations. Typically, one of these concentrations serves as a negative control, i.e. at zero concentration or below the level of detection.

Candidate agents encompass numerous chemical classes, though typically they are organic molecules, preferably small organic compounds having a molecular weight of more than 50 and less than about 2,500 daltons. Candidate agents comprise functional groups necessary for structural interaction with proteins, particularly hydrogen bonding, and typically include at least an amine, carbonyl, hydroxyl or carboxyl group, preferably at least two of the functional chemical groups. The candidate agents often comprise cyclical carbon or heterocyclic structures and/or aromatic or polyaromatic structures substituted with one or more of the above functional groups. Candidate agents are also found among biomolecules including peptides, saccharides, fatty acids, steroids, purines, pyrimidines, derivatives, structural analogs or combinations thereof.

Candidate agents are obtained from a wide variety of sources including libraries of synthetic or natural compounds. For example, numerous means are available for random and directed synthesis of a wide variety of organic compounds and biomolecules, including expression of randomized oligonucleotides and oligopeptides. Alternatively, libraries of natural compounds in the form of bacterial, fungal, plant and animal extracts are available or readily produced. Additionally, natural or synthetically produced libraries and compounds are readily modified through conventional chemical, physical and biochemical means, and may be used to produce combinatorial libraries. Known

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pharmacological agents may be subjected to directed or random chemical modifications, such as acylation, alkylation, esterification, amidification, etc. to produce structural analogs.

Where the screening assay is a binding assay, one or more of the molecules may be joined to a label, where the label can directly or indirectly provide a detectable signal. Various labels include radioisotopes, fluorescers, chemiluminescers, enzymes, specific binding molecules, particles, e.g. magnetic particles, and the like. Specific binding molecules include pairs, such as biotin and streptavidin, digoxin and antidigoxin etc. For the specific binding members, the complementary member would normally be labeled with a molecule that provides for detection, in accordance with known procedures.

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A variety of other reagents may be included in the screening assay. These include reagents like salts, neutral proteins, e.g. albumin, detergents, etc that are used to facilitate optimal protein-protein binding and/or reduce non-specific or background interactions. Reagents that improve the efficiency of the assay, such as protease inhibitors, nuclease inhibitors, anti-microbial agents, etc. may be used. The mixture of components are added in any order that provides for the requisite binding. Incubations are performed at any suitable temperature, typically between 4 and 40°C. Incubation periods are selected for optimum activity, but may also be optimized to facilitate rapid high-throughput screening. Typically between 0.1 and 1 hours will be sufficient.

The compounds having the desired pharmacological activity may be administered in a physiologically acceptable carrier to a host in a variety of ways, orally, topically, parenterally e.g. subcutaneously, intraperitoneally, by viral infection, intravascularly, etc. Depending upon the manner of introduction, the compounds may be formulated in a variety of ways. The concentration of therapeutically active compound in the formulation may vary from about 0.1-100 wt.%. The pharmaceutical compositions can be prepared in various forms, such as granules, tablets, pills, suppositories, capsules, suspensions, salves, lotions and the like. Pharmaceutical grade organic or inorganic carriers and/or diluents suitable for oral and topical use can be used to make up

compositions containing the therapeutically-active compounds. Diluents known to the art include aqueous media, vegetable and animal oils and fats. Stabilizing agents, wetting and emulsifying agents, salts for varying the osmotic pressure or buffers for securing an adequate pH value, and skin penetration enhancers can be used as auxiliary agents.

It is to be understood that this invention is not limited to the particular methodology, protocols, cell lines, animal species or genera, and reagents described, as such may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

As used herein the singular forms "a", "and", and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a cell" includes a plurality of such cells and reference to "the cell" includes reference to one or more cells and equivalents thereof known to those skilled in the art, and so forth. All technical and scientific terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which this invention belongs unless clearly indicated otherwise.

It must be noted that as used herein and in the appended claims, the singular forms "a", "and", and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a complex" includes a plurality of such complexes and reference to "the formulation" includes reference to one or more formulations and equivalents thereof known to those skilled in the art, and so forth.

All publications mentioned herein are incorporated herein by reference for the purpose of describing and disclosing, for example, the methods and methodologies that are described in the publications which might be used in connection with the presently described invention. The publications discussed above and throughout the text are provided solely for their disclosure prior to the filing date of the present application. Nothing herein is to be construed as an

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admission that the inventors are not entitled to antedate such disclosure by virtue of prior invention.

EXPERIMENTAL

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the subject invention, and are not intended to limit the scope of what is regarded as the invention. Efforts have been made to ensure accuracy with respect to the numbers used (e.g. amounts, temperature, concentrations, etc.) but some experimental errors and deviations should be allowed for. Unless otherwise indicated, parts are parts by weight, molecular weight is average molecular weight, temperature is in degrees centigrade; and pressure is at or near atmospheric.

Methods

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Two different types of sequence searches were performed. The first centered on the most highly conserved region of the K+ channel family, the pore domain. The pore is composed of 15-17 amino acids and can be divided into subfamilies based on the number of transmembrane segments present in the channel. Eleven variant peptide sequences corresponding to the pore domain were used in TBLASTN searches against the EST division of Genbank. Significant matches were identified, and classified into 2 categories: identical to known human K+ channels and related to known K+ channels. The pore sequences are shown in Table 2.

TABLE

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SEQ ID NO	Genbank #	
49	L02751 \	TGGTGGGCTGTGGTGACCATGACAACTGTGGGCTATGGGGACATG
50	M60451	TGGTGGGCAGTGGTCACCATGACCACTGTGGGCTACGGGGGACATG
51	L02752	TGGTGGGCAGTCGTCCATGACAACTGTAGGCTATGGAGACATG
52	M55515	TGGTGGGCAGTGGTAACCATGACAACAGTGGGTTACGGCGATATG
53	211585	TGGTGGGCTGTGGTCACCATGACGACCCTGGGCTATGGAGACATG
54	U40990	TGGTGGGGGTGGTCACAGTCACCATCGGCTATGGGGACAAG
55	126643	TGGTGGCCAGTGGTCACCATGACCACGGTTGGCTATGGGGACATG
56	M96747	TGGTGGGCCGTGGTCACCATGACGACCCTGGGCTATGGAGACATG
57	M64676	TGGTGGGCTGTGGTCACCATGACGACACTGGGCTACGGAGACATG
58	M55514	TGGTGGGCTGTGGTGACCATGACAACTGTGGGCTATGGGGACATG
58	X83582	TTCCTGTTCTCCATTGAGACCGAAACAACCATTGGGTATGGCTTCCG
99	578684	TTTTTATTCTCAATAGAGACAGAACCACCATTGGTTATGGCTACCG
19	U22413	TTCCTCTTCTCCATTGAGACCCAGACCATAGGCTATGGTTTCAG
62	U24056	TTCCTGTTCTCGGTGGAGGCGCAGACCATCGGCTATGGGTTCCG
63	U52155	TTCCTCTTCTCCCTTGAATCCCAAACCACCATTGGCTATGGCTTCCG
2	D87281	TITCICITITICCCTGGAATCCCAGACAACCATTGGCTATGGAGTCCG
	D50582	TTCCTTTTCTCCATTGAGGTCCAAGTGACTATTGGCTTTGGGGGGCG
88	D50315	TTTCTCTTCTCCATTGAAGTTCAAGTTACCATTGGGTTTGGAGGGAG
67	U04270	GCGCTCTACTTCACCTTCAGCAGCCTCACCAGTGTGGGCTTCGGCAAC

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The unique pore peptides sequences are shown in Table 3.

TABLE 3

IADLE 3
Amino acid Sequence
WWAVVSMTTVGYGDM
WWAVVTMTTLGYGDM
WWGVVTVTTIGYGDK
WWAVVTMTTVGYGDM
FLFSIEVQVTIGFGG
FLFSLESQTTIGYGV
FLFSIETETTIGYGY
FLFSIETQTTIGYGF
FLFSVETQTTIGYGF
FLFSLESQTTIGYGF
FLFSIETETTIGYGF
ALYFTFSSLTSVGFGN

The second set of experiments was based on a complex, reiterative process.

Annotated protein and DNA sequences were obtained from GenBank for all known K+ channels from all species. The TBLASTN and BLASTN programs were used to identify homologous ESTs, which were then analyzed using the BLASTX and BLASTN algorithms to identify ESTs which were related to K+ channels yet not identical to any known human K+ channel gene.

Novel human K+ channels were defined as those that had clear homology to known K+ channels from any species and were not present as identities or near identities to any human-derived sequences in any division of Genbank.

Isolation of full length cDNA sequence. EST clones were picked from the IMAGE consortium cDNA library and end-sequenced with vector primers. Gap closure was achieved either by primer walking or transposon sequencing. GeneTrapper (Life

Technologies) was used to isolate larger cDNA clones according to the provided protocol. RACE was used to extend the sequences as necessary using standard protocols.

Sequences were assembled in Sequencher (Gene Codes). The presence of open reading frames was assessed as well as potential start codons. Potential polymorphisms were detected as sequence variants between multiple independent clones. Sequence homologies were detected using the BLAST algorithms.

The completed gene sequences and predicted amino acid sequences are provided as SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21-24, 26 and 28-29. Polymorphisms, chromosome locations and family assignments are shown in Table 1.

ESTs that had top human hits with >95% identity over 100 amino acids were discarded. This was based upon the inventors' experience that these sequences were usually identical to the starting probe sequences, with the differences due to sequence error. The remaining BLASTN and BLASTX outputs for each EST were examined manually, i.e., ESTs were removed from the analysis if the inventors determined that the variation from the known related probe sequence was a result of poor database sequence. Poor database sequence was usually identified as a number of 'N' nucleotides in the database sequence for a BLASTN search and as a base deletion or insertion in the database sequence, resulting in a peptide frameshift, for a BLASTX output. ESTs for which the highest scoring match was to non-related sequences were also discarded at this stage. The EST sequences that correspond to each clone are shown in Table 4.

Table 4 January 1997 St. A. A. A. A.

Genbank Accession#			Trace : "	IMAGE Plate Coordinates	Read 5'/3'
N39619	K+Hnov2	277113	yy51h05.s1	611p10	3'
N46767	K+Hnov2	277113	yy51h05.r1	611p10	5'
R19352	K+Hnov11	33144	yg24f12.r1	155024	5'
R44628	K+Hnov11	33144	yg24f12.s1	155024	3'

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R35526	K+Hnov14	37299	yg64e08.r1	165015	5'
R73353	K+Hnov14	157854	yl10e04.r1	7, 251g07 (3) (4) (4)	5'
AA397616	K+Hnov14	728558	zt79c08.r1	1787j15	5'
AA286692	K+Hnov28	700757	zs48h03.r1	1715d6	5'
AA150494	K+Hnov42	491748	zl08e07.s1	1170013	3'
AA156697	K+Hnov42	491748	zl08e07.r1	1170013	5'
AA191752	K+Hnov42	626699	zp82d06.r1	1522i12	5'
AA216446	K+Hnov42	626699	zp82d06.s1	1522f12	3'
AA430591	K+Hnov42	773611	zw51f10.r1	1904020	5'
AA236930	K+Hnov44	683888	zs01a05.s1	1671e9	3'
AA236968 `	K+Hnov44	683888	zs01a05.r1	1671e9	5'

EXAMPLE 2: CHROMOSOMAL LOCALIZATION

Two primers were designed in the 3'-untranslated regions of each gene sequence to amplify a product across the Stanford G3 radiation hybrid map, or the Whitehead GB4 panel. The PCR data were submitted for automatic two-point analysis. Mapping data were correlated with cytoband information and comparisons with the OMIM human gene map data base were made. The following primers were made:

K+Hnov1 on GB4 (SEQ ID NO:31) F: 5' TATCCACATCAATGGACAAGC 3' (SEQ ID NO:32) R: 5' TGCATAACTGGCTGGGTGTA 3' Results: 1.71 cR from D2S331, Cytogenetic location of 2g37

K+Hnov2 on G3
15 F: 5' GTCAGGTGACCGAGTTCA 3'
R: 5' GCTCCATCTCCAGATTCTTC 3'
Results: 0.0 cR from SHGC-1320, Cytogenetic location of 11q12

K+Hnov6 on GB4

20 (SEQ ID NO:33) F: 5' TGACATCACTGGATGAACTTGA 3'
(SEQ ID NO:34) R: 5' TGCCTGCAAAGTTTGAACAT 3'
Results: 5.23 cR from WI-5509, Cytogenetic location of 2p23

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K+Hnov9 on GB4
25 (SEQ ID NO:35) F: 5' TGACATCACTGGATGAACTTGA 3' (SEQ ID NO:36) R: 5' TGCCTGCAAAGTTTGAACAT 3'

Results: 1.21 cR from AFM200VC7, Cytogenetic location of 8q23

K+Hnov11 on GB4

(SEQ ID NO:37) F: 5' ACCTGGTGGTATGGAAGCAT 3'

5 (SEQ ID NO:38) R: 5' TTTCTCCTGGCCTCTACCC 3'

Results: 2.43 cR from WI-6756, Cytogenetic location of 8q23

K+Hnov12 on G3

(SEQ ID NO:39) F: 5' TCCCTCTTGGGTGACCTTC 3'

(SEQ ID NO:40) R: 5' ATCTTTGTCAGCCACCAGCT 3' 10

Results: 7.45 cR from SHGC-32925, Cytogenetic location of Xp21

K+Hnov14 on GB4

(SEQ ID NO:41) F: 5' AGGTGTGCTGCCATCTGCTGTTCG3'

15 (SEQ ID NO:42) R: 5' AGCCTATCCTCTGAGAGTCAGG

Results:

7.69 cR from WI-7107, Cytogenetic location of 12q14

K+Hnov28 on GB4

(SEQ ID NO:43) F: 5' AAGCAGAGTACTCATGATGCC 3'

20 (SEQ ID NO:44) R: 5' TCTGGTAGACAGTACAGTGG 3'

Results: 35.38 cR from WI-9695, Cytogenetic location of 3q29

K+Hnov42 on G3

(SEQ ID NO:45) F: 5' CATTTGGCTGGTCCAAGATG 3' Common Common

(SEQ ID NO:46) R: 5' AGTCATTGGTAGGGAGGTAC 3' 25

Results: 7.45 cR from SHGC-32925, Cytogenetic location of Xp21

K+Hnov44 on G3

(SEQ ID NO:47) F: 5' CATGCTTCTACAGTCCAGCC 3'

30 (SEQ ID NO:48) R: 5' GGTCCTCAGTTGCAGAAATC 3'

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Results: 7.45 cR from SHGC-32925, Cytogenetic location of Xp21

Map positions for K+Hnov15 and K+Hnov27 were obtained from public databases. K+Hnov2 and K+Hnov4 have not been mapped.

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EXAMPLE 3: EXPRESSION ANALYSIS

RT-PCR was utilized to characterize the expression pattern of the novel ion channels. This approach used RNA from 30 different tissues to generate first strand cDNA. Total RNA was purchased (Clontech, Invitrogen) and used to synthesize first strand cDNA using M-MLV reverse transcriptase and the supplied buffer (Gibco-BRL). The 20 µl reaction contained 5 µg total RNA, 100 ng of random primers, 10 mM DTT.

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0.5 mM each dNTP, and an RNAse inhibitor (Gibco-BRL). Identical reactions were set up without reverse transcriptase to control for DNA contamination in the RNA samples. The synthesis reaction proceeded for 1 hour at 37°C followed by 10 minutes at 95°C. These cDNAs, along with control cDNA synthesis reactions without reverse transcriptase, were diluted 1.5 and 2 μl of each sample were arrayed into 96-well trays, dried, and resuspended in PCR buffer prior to PCR amplification. The cDNAs were tested with primers with defined expression patterns to verify the presence of amplifiable cDNA from each tissue. Gene-specific primers were used to amplify the cDNAs in 20 μl PCR reactions with standard conditions, 2.5 mM MgCl₂, Taq Gold, and an appropriate annealing temperature.

This approach provides for relatively high-throughput analysis of gene expression in a large set of tissues in a cost-efficient manner and provides qualitative analysis of gene expression only. Modifications can be employed, such as the use of internal control primers, limited cycling parameters, and dilution series to convert this to a quantitative experiment.

Table 3

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Trachea	٠			•	•	•	\cdot	٠	٠	٠	•		•
Thymus	•	٠		•	•	7	$\overline{\cdot}$	•	1	•	•	1	•
Testis	•	•		•	•	\exists	-	•	•	•		7	7
Stomach		-	-	-			7		•	٠	1	+	
Spleen	•	-			-	┪		1	-	•		┪	\dashv
Smail	•			-	4		_	┵	4	-	Ĭ	\dashv	-
Intestine Skin		•	•	•	4	-	-	5.,	1	•	-	4	-
Skeletal				-	-	_		•	4	•	4		_
Muscle 🚉 👢	٠	*	٠	٠	_	_		•		•	•	4	•
Salivary Gland	٠	\Box	٠	٠	<u>•</u>	٠	•	•	1	٠	٠		٠
Rectum		٠		٠	·		\cdot	•	ا	٠			٠
Prostate	٠	•	•	•		٠		٠	٠	٠	•		•
Placenta	•	•	·	•	•	٠	•	٠	٠	٠	٠		٠
Pancreas 377 7	٠	•	•	٠	•	•		٠	٠	٠	٠		•
Mammary Gland	•	•	•	٠	•	•	•	٠	•	٠	٠		•
Lung	•	•	•	•		٠	•	٠	٠	+	•		•
Liver	•	•	•	•		•	•	•	-	•	•		•
Kidne.	•	•	-	•	\vdash			•		•			•
HeLa Co		Ľ		·	_	•	•	•	•	_		Н	•
Heart :	H	Ļ				Ž			_	_	_		
Fetal Late	ŀ	•	Ŀ	•	•	•	٠	•	•	•	•		•
	•	•	Ŀ	·	٠	•	•	•	•	•	•		٠
Fetal Er :	Ŀ	Ŀ	٠	٠	\cdot	٠	٠	٠	٠	٠	•		•
Esopran	٠	Ŀ	L	•			\cdot	٠		*	٠		٠
Cej.cn	٠	٠	٠	٠	$oxed{\cdot}$	$\lfloor \cdot floor$		·	•	•	٠		٠
Cervik .	·	٠					•	•		•			•
Cerebo um	+	•	٠	٠	•	•	•	٠	+	•	•	П	•
Brain	•	•	٠	٠	•	•	•	.	•	•	•	П	•
Blaccer	•	•		•		•	•	•	•	•	•	П	•
»Àdrenat	-	╁		-				•	•	•		۲	
Gland Adipose	ŀ	-	\vdash	+	+	H	-	-	-	-			-
Anchor name	Ė	F	\vdash	\vdash	150			Ė			ŀ		É
	_	2	-	9	æ	Ę	7	=	15	27	8	Ş	Ŧ
	K. Hnov1	K+Hnov2	K+Hnov4	+Hnov6	.Hnov9	K+Hnov1	K+Hnov12	+Hnov14	+Hnov15	K+Hnov27	K+Hnov28	K+Hnov42	K+Hnov44
	ş	\$	<u>₹</u>	3	¥	¥	¥	혹	¥	ż	¥	¥	¥

A *+* indicates expression in the tissue, a *-* indicates no expression, and blank square indicates no data for that sample.

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K+Hnov49 on Whitehead GB4 RH mapping panel:

Primer 1 (SEQ ID NO:5): 5' - CATAGCCATAGGTGAGGACT - 3'

Primer 2: (SEQ ID N:6) 5" - GAGAGGAAAACAGTCTGGGC - 3"

5 Results: Cytogenetic location 1q41, 4.6cR from framework marker D1S217

K+Hnov59 on Whitehead GB4 RH mapping panel

Primer 1 (SEQ ID NO:7): 5' - GGACATCGAACTAAGACCTG - 3'

Primer 2 (SEQ ID NO:8): 5' - TCCCATGCCATTCAGATCTG - 3'

10 Results: Cytogenetic location 19q13.2, 8.34cr from framework marker D19S425

EXPRESSION ANALYSIS OF K+HNOV49

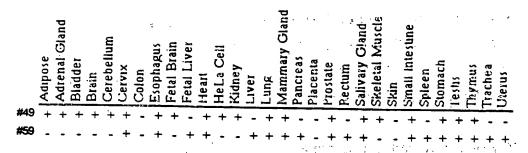
A probe was created from a fragment corresponding to nucleotides 50 to 1284 of SEQ ID NO:83 (K+Hnov49) and purified DNA fragment was labeled with [³²P]dCTP (Amersham) by the random primer method. Adult human Multiple Tissue Northern (MTM™) Blots (Clontech) were hybridized with the [³²P]-labeled fragment in ExpressHyb™ solution (Clontech) for four hours, washed to a final stringency of 0.1xSSC, 0.1% SDS at 65°C and subjected to autoradiography for 24 hours.

Analysis revealed that K+Hnov49 is expressed as an approximately 4.2kb mRNA. Expression levels of K+Hnov49 are high in brain and liver and low in kidney tissues. No mRNA was detectable on these Northern blots for heart, skeletal muscle, colon, thymus, spleen, small intestine, placenta, lung or peripheral blood leukocytes indicating either a very low level of expression or that it is not expressed in these tissues. Expression analysis was also carried out by RT-PCR across an extended series of tissues. The results of these analyses are shown in Table 4. Primer pairs used for amplification of K+Hnov49 and 59 are the same as those used for RH mapping as indicated above.

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WHAT IS CLAIMED IS:

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- 1. An isolated nucleic acid encoding a mammalian K+Hnov protein.
- 2. An isolated nucleic acid according to Claim 1, wherein said K+Hnov protein has the amino acid sequence of SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 25, 27, 30, 81 or 83.
- 3. An isolated nucleic acid according to Claim 1, wherein said K+Hnov protein has an amino acid sequence that is substantially identical to the amino acid sequence of SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 25, 27, 30, 81 or 83.
- 4. An isolated nucleic acid according to Claim 1 wherein the nucleotide sequence of said nucleic acid is SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 22, 23, 24, 26, 28, 29, 80 or 82.
 - 5. An isolated nucleic acid that hybridizes under stringent conditions to a nucleic acid sequence of claim 4.
 - 6. An expression cassette comprising a transcriptional initiation region functional in an expression host, a nucleic acid having a sequence of the isolated nucleic acid according to Claim 1 under the transcriptional regulation of said transcriptional initiation region, and a transcriptional termination region functional in said expression host.

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7. A cell comprising an expression cassette according to Claim 6 as part of an extrachromosomal element or integrated into the genome of a host cell as a result of introduction of said expression cassette into said host cell, and the cellular progeny of said host cell.

8. A method for producing mammalian K+Hnov protein, said method comprising:

growing a cell according to Claim 7, whereby said mammalian K+Hnov protein is expressed; and

isolating said K+Hnov protein free of other proteins.

- 9. A purified polypeptide composition comprising at least 50 weight % of the protein present as a K+Hnov protein or a fragment thereof.
- 10. A monoclonal antibody binding specifically to a K+Hnov protein.
 - 11. A non-human transgenic animal model for K+Hnov gene function wherein said transgenic animal comprises an introduced alteration in a K+Hnov gene.

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- 12. The animal model of claim 11, wherein said animal is heterozygous for said introduced alteration.
- 13. The animal model of claim 12, wherein said animal is homozygous20 for said introduced alteration.
 - 14. The animal model of claim 12, wherein said introduced alteration is a knockout of endogenous K+Hnov gene expression.

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SEQUENCE LISTING

<110> Miller, Andrew Curran, Mark Buckler, Alan <120> Novel Human Potassium Channels <130> SEQ-15PCT <150> 60/076,687 <151> 1998-02-25 <150> 60/095,836 <151> 1998-08-07 <150> 60/116,448 <151> 1999-01-19 <170> FastSEQ for Windows Version 3.0 210> 1 <211> 2932 <212> DNA <220> <221> CDS <222> (103)...(1180) <223> K+Hnov1 attaaaatta tetgateaaa aaggeagaet etgtaaattt eettaagaee taeettggea 60 taaaggetga eecageaaaa gaactgagaa atacageetg ag atg gae age agt 114 Met Asp Ser Ser **1** 1 aat tgc aaa gtt att gct cct ctc cta agt caa aga tac cgg agg atg 162 Asn Cys Lys Val Ile Ala Pro Leu Leu Ser Gln Arg Tyr Arg Arg Met 15 gtc acc aag gat ggc cac agc aca ctt caa atg gat ggc gct caa aga 210 Val Thr Lys Asp Gly His Ser Thr Leu Gln Met Asp Gly Ala Gln Arg 25 ggt ctt gca tat ctt cga gat gct tgg gga atc cta atg gac atg cgc 258 Gly Leu Ala Tyr Leu Arg Asp Ala Trp Gly Ile Leu Met Asp Met Arg 45 tgg cgt tgg atg atg ttg gtc ttt tct gct tct ttt gtt gtc cac tgg 306 Trp Arg Trp Met Met Leu Val Phe Ser Ala Ser Phe Val Val His Trp 60 (65 ctt ytc ttt gca gtg ctc tgg tat gtt ctg gct gag atg aat ggt gat Leu Val Phe Ala Val Leu Trp Tyr Val Leu Ala Glu Met Asn Gly Asp 75 80

ctg Leu 85	gaa Glu	cta Leu	gat Asp	cat His	gat Asp 90	gcc Ala	cca Pro	cct Pro	gaa Glu	aac Asn 95	cac	act Thr	atc Ile	tgt Cys	gtc Val 100	402
aag Lys	tat Tyr	atc Ile	acc Thr	agt Ser 105	ttc Phe	aca Thr	gct Ala	gca Ala	ttc Phe 110	tcc Ser	ttc Phe	tcc Ser	ctg Leu	gag Glu 115	aca Thr	450
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agt Ser	gca Ala	atc Ile 135	gcc Ala	tta Leu	ctt Leu	gcc Ala	ata Ile 140	caa Gln	atg Met	ctc Leu	cta Leu	ggc Gly 145	ctc Leu	atg Met	cta Leu	546
gag Glu	gct Ala 150	ttt Phe	atc Ile	aca Thr	ggt Gly	gct Ala 155	ttt Phe	gtg Val	gcg Ala	Lys	att Ile 160	gcc Ala	cgg Arg	cca Pro	aaa Lys	594
Asn 165	Arg	Ala	Phe	Ser	Ile 170	Arg	Phe	Thr	Asp	Thr 175	Ala	Val	gta Val	Ala	His 180	642
atg Met	gat Asp	ggc	aaa Lys	cct Pro 185	aat Asn	ctt Leu	atc Ile	ttc Phe	caa	gtg	gcc	aac	acc Thr	cga Arg 195	cct Pro	690
Ser	Pro	Leu	Thr 200	Ser	Val	Arg	Val	Ser 205	Ala	Val	Leu	Tyr	cag Gln 210	gaa Glu	Arg	738
gaa Glu	aat Asn	ggc Gly 215	aaa Lys	ctc Leu	tac Tyr	cag Gln	acc Thr 220	agt Ser	gtg Val	gat Asp	ttc Phe	cac His 225	ctt Leu	gat Asp	Gly	786
Ile	Ser 230	Ser	Asp	Glu	Cys	Pro 235	Phe	Phe	Ile	Phe	Pro 240	Leu	Thr.	Tyr.	tat Tyr	834
cac His 245	tcc Ser	Ile	aca Thr	cca Pro	ser 250	agt Ser	cct Pro	ctg Leu	gct Ala	act Thr 255	Leu	ctc Leu	cag Gln	cat His	gaa Glu 260	882
aat Asn	Pro	tct Ser	His	ttt Phe 265	gaa Glu	tta Leu	gtt Val	gta Val	ttc Phe 270	ctt Leu	tca Ser	gca Ala	atg Met	cag Gln 275	gag Glu	930
ggc	act Thr	gga Gly	gaa Glu 280	ata Ile	tgc Cys	caa Gln	agg Arg	agg Arg 285	aca Thr	tcc Ser	tac Tyr	cta Leu	ecg Pro 290	tct Ser	gaa Glu	978
atc Ile	atg Met	tta Leu 295	His	cac His	tgt Cys	ttt Phe	gca Ala 300	tct Ser	Leu.	Leu.	acc Thr	Arg		tcc Ser	aaa Lys	1026
ggt Gly	gaa Glu 310	tat Tyr	caa Gln	Ile	aag Lys	atg Met 315	gag Glu	aat Asn	ttt Phe	qaA	aag Lys 320	act Thr		cct Pro	Glu	1074
ttt	cca	act	cct	ctg	gtt	tct	aaa	agc	cca	aac	agg	act	gac	ctg	gat	1122

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          330
                                        335 340
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                                                                   1170
 Ile His Ile Asn Gly Gln Ser Ile Asp Asn Phe Gln Ile Ser Glu Thr
                345
                                    350
 gga ctg aca g aataagactt atccattttt taatgtatta aatacaccca
                                                                   1220
 Gly Leu Thr
 gecagitatg cagetactit tietttactg tateteatgt titettitt caatgetaat
                                                                   1280
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                                                                   1340
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caaaacaaaa accaaaaaag tcaccctatc aggtttcaaa cagatttgtg gctgttcttt
                                                                   2480
 totgaaattt coottattca ggtttotgtg ggaaaaatga aagattaaco ttocccactg
                                                                   2540
 gtgatgacct aggcaggaat catctcttga aataaatact agctgagtaa aggcaagcag
                                                                   2600
 gtgtgaagag cagggctcag cagcaagtca catttttcta ctatttgacc aaaaggaaaa
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 Val Val His Trp Leu Val Phe Ala Val Leu Trp Tyr Val Leu Ala Glu
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 Met Asn Gly Asp Leu Glu Leu Asp His Asp Ala Pro Pro Glu Asn His
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 Thr Ile Cys Val Lys Tyr Ile Thr Ser Phe Thr Ala Ala Phe Ser Phe
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 Ala Arg Pro Lys Asn Arg Ala Phe Ser Ile Arg Phe Thr Asp Thr Ala
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 Val Val Ala His Met Asp Gly Lys Pro Asn Leu Ile Phe Gin Val Ala
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 Asn Thr Arg Pro Ser Pro Leu Thr Ser Val Arg Val Ser Ala Val Leu
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 Ala Met Gln Glu Gly Thr Gly Glu Ile Cys Gln Arg Arg Thr Ser Tyr
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 Arg Gly Ser Lys Gly Glu Tyr Gln Ile Lys Met Glu Asn Phe Asp Lys 305 310 315
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                                                         164
 Glu Ala Ser Glu Lys Ile Ile Ile Asn Val Gly Gly Thr Arg His Glu
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 Thr Tyr Arg Ser Thr Leu Arg Thr Leu Pro Gly Thr Arg Leu Ala Trp
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ggc Gly 85	aag Lys	ctg Leu	cac His	tgc Cys	ccc Pro 90	gca Ala	gac Asp	gtg Val	tgc Cys	ggg Gly 95	ccg Pro	ctc Leu	ttc Phe	gag Glu	gag Glu 100	404
Glu	ren	Ala	Phe	Trp 105	Gly	acc Ile	Asp	Glu	Thr 110	Asp	Val	Glu	Pro	Cys 115	Суз	452
Trp	Met	rnr	120	Arg	Gln	cac His	Arg	Asp 125	Ala	Glu	Glu	Ala	Leu 130	Asp	Ile	500
Pne	GIU	135	Pro	Asp	Leu	att Ile	Gly 140	Gly	Asp	Pro	Gly	Asp 145	Asp	Glu	Asp	548
Leu	150	Aia	Lys	Arg	Leu	ggc Gly 155	Ile	Glu	Asp	Ala	Ala 160	Gly	Leu	Gly	Gly	596
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Phe	Leu	Val	Arg	11e 265	Val	ttt Phe	Ser	Pro	Asn 270	Lys	Leu	Glu	Phe	Ile 275	Lys	932
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Glu	Val	Gly 295	Leu	Ser	Gly	Leu	Ser 300	Ser	Lys :	Ala	Ala	Lys 305	Asp	Val	Leu	
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ctt Leu	ccá Pro	agg Arg 455	aaa Lys	aga Arg	aag Lys	aag Lys	cac His 460	atc Ile	cct Pro	cct Pro	gct Ala	cct Pro 465	cag Gln	gca Ala	agc Ser	1508
Ser	cct Pro 470	act Thr	ttt Phe	tgc Cys	aag Lys	aca Thr 475	gaa Glu	tta Leu	Asn	atg Met	gcc Ala 480	tgc Cys	aat Asn	agt Ser	aca Thr	1556
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Gln 485 aga	Ser	Asp	Thr	Cys	Leu	Gly gac	Lys	Asp agt	aat Asn aca	Arg 495 gga	ctt Leu agt	Leu	Glu	His cca	Asn 500 cta Leu	1604
Gln 485 aga Arg	ser tca ser	Asp gtg Val cca	Thr tta Leu gaa	tca Ser 505	Leu 490 ggt	gac Asp	Lys gac Asp	agt Ser	aat Asn aca Thr 510	Arg 495 gga Gly tct	ctt Leu agt Ser	Leu gag Glu acc	Glu ccg Pro	cca Pro 515	Asn 500 cta Leu	

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Cys Ala Ser 550	Asp Gly Gl	/ Ile Arg Lys 555	Gly Tyr Glu	Lys Ser Arg	y Ser
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Asp GIY GIY	Gly Val Gly	Ser Ser Gly 55	Ser Ser Gly	Gly Gly Gly	Cys
Glu Phe Phe	Phe Asp Arg		Val Phe Ala	Tyr Val Leu	Asn
65 Tyr Tyr Arg	70 Thr Glv Lvs	Leu His Cvs	75 Pro Ala Asp	Val Cue Cla	80
•	85	,	90	• 95	-
Leu Phe Glu	Glu Glu Leu 100	Ala Phe Trp 105	Gly Ile Asp	Glu Thr Asp	Val
Glu Pro Cys			Gln His Arg	Asp Ala Glu	Glu
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Gly Leu Gly			Gly Arg Trp	Arg Arg Leu	160 Gln
Pro Arg Met	165 Tro Ala Leu	Phe Glu Aco	170 Pro Tyr Ser	175	21-
	180	185		190	
Arg Phe Ile	Ala Phe Ala	Ser Leu Phe 200	Phe Ile Leu		Thr
Thr Phe Cys	Leu Glu Thr		Phe Asn Ile	205 Val Lys Asn	Lys
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225	230		Val Val Leu 235	1 1 2 1	240
Glu Thr Asp	Pro Ala Leu	Thr Tyr Val	Glu Gly Val	Cys Val Val	Trp
Phe Thr Phe	245 Glu Phe Leu	Val Arg Ile	250 Val Phe Ser	Pro Asn Lve	Leu
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Giu Phe Ile 275	Lys Asn Leu	Leu Asn Ile 280	Ile Asp Phe	Val Ala Ile	Leu

285

280

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Lys Ser Arg Ser Leu Asn Asn Ile Ala Gly Leu Ala Gly Asn Ala Leu
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Met Val Phe Gly Glu Phe Phe His
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aag Lys 25	cag Gln	tct Ser	gtt Val	Asp	caa Gln 30	agc Ser	acc Thr	ctc Leu	ctg Leu	cgg Arg 35	ttt Phe	cct Pro	cac His	acc	aga Arg 40	449
ctg Leu	Gly 999	aag Lys	ctg Leu	ctt Leu 45	act Thr	tgc Cys	cat His	tct Ser	gaa Glu 50	gag Glu	gcc Ala	att	ctg Leu	gag Glu 55	ctg Leu	497
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tgt Cys	gat Asp	gat Asp	tac Tyr 60	agt Ser	gtg Val	gcc Ala	gat Asp	aag Lys 65	gaa Glu	tac Tyr	tac Tyr	ttt Phe	gat Asp 70	cgg Arg	aat Asn	545
2										Marian Control	,		e 1		•	
Pro	Ser	Leu 75	Phe	aga Arg	tat Tyr	gtt Val	Leu 80	aat Asn	Phe	tat Tyr	tac Tyr	acg Thr 85	Gly ggg	aag Lys	ctg Leu	593
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His	Val 90	Met	Glu	Glu	Leu	Cys 95	Val	ttc Phe	Ser	Phe	tgc Cys 100	Gln	gag Glu	Ile	gag Glu	641
+20	+~~					- 22 -	1									
Lac	cgg	ggc	atc	aac	gag	CEC	TTC	att	gat	tct	tgc	tgc	agc	aat	cgc	689
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105					110		**			115	;	٠.			120	
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com	Ude	300	9-9	Com	mb-	gac	200	cog	27	yaa	gag	rcg	- CC	ctg	ככנ	785
Ser	uis	ASP		ser	Thr	Asp	ser	Ser	Pne	Glu	Glu	Ser	Ser	Leu	Phe	•
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Glu	Lys	Glu	Leu	Glu	Lys	Phe	Asp	Thr	Leu	Arg	Phe	Gly	Gln	Leu	Arg	
	_	155			-		160			_		165				
	•							٠.								
aaσ	aaa	atc	taa	att	aga	ato	gag	aat	cca	aca	T 3.0	+~~	a+a	+00	~~+	881
Lvs	Lvs	Tle	Trn	Tle	Ara	Mer	Glu	Asn	Dro.	772	The	Cura	Tan	C	21-	
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Ala	Met	Сув	Val	His	Ser	Met	Ser	Glu	Phe	Gln	Asn	Glu	Asp	GIV	G] 11	/
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	· :				٠.						·:		•		_	
ara	cra+	ra+		ata	cta	<i>(</i> -	- · · ·	~+~	~ : ~ : ~ :	-+-		*			-	1005
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val	Asp	qsa			ren	GIU	GTĀ	Val	GIU	TTE	Ala	Сув			Trp	
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Phe	Thr	Gly	Glu	Leu	Ala	Val	Arg	Leu	Ala	Ala	Ala	Pro	Cys	Gln	Lys	
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490

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Thr Thr Val Gly Tyr Gly Asp Thr His Pro Val Thr Leu Ala Gly Lys
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Pro Ile Thr Ile Ile Phe Asn Lys Phe Ser Lys Tyr Tyr Gln Lys Gln
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Lys Asp Ile Asp Val Asp Gln Cys Ser Glu Asp Ala Pro Glu Lys Cys
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His Glu Leu Pro Tyr Phe Asn Ile Arg Asp Ile Tyr Ala Gln Arg Met
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His Ala Phe Ile Thr Ser Leu Ser Ser Val Gly Ile Val Val Ser Asp
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Glu Pro Leu Ala Leu Gly Asp Cys Phe Thr Val Asn Val Gly Gly Ser
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Arg Phe Val Leu Ser Gln Gln Ala Leu Ser Cys Phe Pro His Thr Arg 50 55 60
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Leu Gly Lys Leu Ala Val Val Val Ala Ser Tyr Arg Arg Pro Gly Ala
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ctg gcc gcc gtg ccc age cct ctg gag ctt tgc gac gat gcc aac ccc
Leu Ala Ala Val Pro Ser Pro Leu Glu Leu Cys Asp Asp Ala Asn Pro
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150,000

 $(x,y) \leq (\phi_0)$

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tgc Cys	gcg Ala 130	ctc Leu	tcc Ser	ttc Phe	ctg Leu	cag Gln 135	gag Glu	atc Ile	cag Gln	tac Tyr	tgg Trp 140	ggc	atc Ile	gat Asp	gag Glu	911
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		•														
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Gln	Ser	Ile	Pro	Asp	Thr	Thr	Phe	Thr	Ser	Val	Pro	Cys	Ala	Tro	Tro	
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	•													•	•	
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-					• •					•				413	, .	
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Ile	Leu	Val	Leu	Ala	Leu	Pro	Tle	Ala	Tle	Tile	Age	700	7	Dha	Com	1//5
2	٠.		420					425			<i>79</i> 11	. vob.	430	File	261	
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GIU	450	пеп	TAR	riAs	reu		гăа	Asn.	TTE	Ala		Asp	Ser	Tyr	Ile	
	450				P .	455					460		•	-		
act	arr	226	++~	363	~a+						- 4					
Sar	Val	Aco	Lou	aya N==	gac 3an	y.c.	Cat	gcc	cgg	agt	atc	atg	gag	atg	ctg	1919
465	Val	və.i.	Den	Arg	ASD	Val.	TYE	Ala			T.ī.e	Met	GIA			
405		:		•	470	•	•	٠.		475		٠.			480	
CCS	cta		~~~	202	~					:						
y z z	Lou	Tues	23	aya N==	gaa	aya	gca	agt	act	agg	agc	agc	a aa	gga	gat	1967
ALG	pen	Lys	GLY		GIU	Arg	ALA .	Ser		Arg	Ser	Ser	Gly		Asp	*
	• . •		· .	485	•		•		490				•	495	***	
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yat Nam	Dho	rgg	ב בכ	tgaa	ttaa	בבב	tcaa	ttt	attt	acaa	aa g	ctat	gtac	a		2017
Asp	PHE	тър	. ^		•				• •	?	: :					
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a++-									.							
alta	acta	aa a	-yat	aaag	c ag	rgat	9 - 99	att	cctg	tat	tctg	atga	tg a	gtct	cttca	2077
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135
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Ile Leu Val Leu Ala Leu Pro Ile Ala Ile Ile Asn Asp Arg Phe Ser

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	450				Leu	455					460	•		_		
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Asp	Phe	Trp				-			7,00	• • •				422	÷*	
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CCC	geet	tot (taga	acac	2C C	cagti	ctcct	t tgt	ttg	ggtg	get	caag	gtg	tctc	caagco	180
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Tyr	Pro	Asp	Thr	Leu	Leu	Gly	Ser	Ser	Glu	Lys	Glu	Phe	Phe	Tvr	gat Asp	484
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His	Val	Leu	Asn	Phe	Tyr	Arg	Thr	Gly	Arg	Leu	His	Cys	Pro	Arg	Gln	
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ansidulià i ultilatea di

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EDC 117

9840

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645

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ccg Pro	gcc Ala	agg Arg 55	gct' Ala	gcg Ala	ctg Leu	ctg Leu	cag Gln 60	gca Ala	gtt Val	gca Ala	ctg Leu	gga Gly 65	ctg Leu	ctg Leu	gtg Val	368
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cac His	ccc Pro	gtg Val	att Ile 120	tac Tyr	Cac	ctg Leu	Gly	cag Gln 125	Leu	Ala	ctt Leu	ctt Leu	ggt Gly 130	tac Tyr	ttg Leu	560
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				5 Pro	Ala	Ser	Leu	Ala 25	10 Leu	Val-	Ala	Thr	Leu 30	15 Arg	His	
Cys	Leu	Leu	-	Val	Leu	Ser	Arg		Arg	Ala	Trp	Val		Val	His	

0 / 10 /WO__ 6 /436-641_1 >

Try Gin Leu Ser Pro Ala Arg Ala Ala Leu Leu Gin Ala Val Ala Leu 50 5 5 6 60 70 70 70 80 75			35					40					4 =			•	
Gly Leu Leu Val Ala Ser Ser Phe Val Leu Leu Pro Ala Leu Val Leu 65 70 Trp Gly Leu Gln Gly Asp Cys Ser Leu Leu Gly Ala Val Tyr Phe Cys 85 90 Phe Ser Ser Leu Ser Thr Ile Gly Leu Glu Asp Leu Leu Pro Gly Arg 100 105 110 Gly Arg Ser Leu His Pro Val Ile Tyr His Leu Gly Gln Leu Ala Leu 115 Leu Gly Tyr Leu Leu Leu Gly Leu Leu Ala Met Leu Leu Ala Val Glu 130 Thr Phe Ser Glu Leu Pro Gln Val Arg Ala Met Gly Lys Phe Phe Arg 145 Thr Phe Ser Gly Pro Val Thr Ala Glu Asp Gln Gly Gly Ile Leu Gly Gln 165 Asp Glu Leu Ala Leu ser Thr Leu Pro Pro Ala Ala Pro Ala Ser Gly 180 Gln Ala Pro Ala Cys 190 Gln Ala Pro Ala Cys 190 <pre></pre>	Trp	Gln 50	Leu	Ser	Pro	Ala	Arg	Ala	Ala	Leu	Leu		45 Ala	Val	Ala	Leu	
Trp Gly Leu Gln Gly Asp Cys Ser Leu Leu Gly Ala Val Tyr Phe Cys 85 90 Phe Ser Ser Leu Ser Thr Ile Gly Leu Glu Asp Leu Leu Pro Gly Arg 100 115 110 115 120 120 121 120 121 Leu Gly Tyr Leu Leu Leu Gly Leu Leu Ala Met Leu Ala Leu 130 Thr Phe Ser Glu Leu Pro Gln Val Arg Ala Met Gly Lys Phe Phe Arg 145 Pro Ser Gly Pro Val Thr Ala Glu Asp Gln Gly Gly Ile Leu Gly Gln 185 Pro Ser Gly Pro Val Thr Ala Glu Asp Gln Gly Gly Ile Leu Gly Gln 185 Asp Glu Leu Ala Leu Ser Thr Leu Pro Pro Ala Ala Pro Ala Ser Gly 180 Gln Ala Pro Ala Cys 195 Gln Ala Pro Ala Cys 195 <210> 17 <221> 3102 <221> DNA <2213> H. sapiens <222> (274) (1705) <222> (274) (1705) <222> (274) (1705) <222> (274) (1705) <223> K+Hnov11 <400> 17 gcacgcgcaa agcgccaacc ggagaccctg gggtggagct tgtggtaata gaaacatacc caccccaga ctttcctggg aggggacccg ggcgccag taatgggtag ggagaggggg cccccccccagc ctttcctggg aggggacccccc aggacccacc aggaggcccccc aggacccacc ag at ac ac gcc ggc gcccccccccc	Gly 65	Leu	Leu	Val	Ala	Ser	Ser	Phe	Val	Leu	Leu	Pro	Ala				
Phe Ser Ser Leu Ser Thr Ile Gly Leu Glu Asp Leu Leu Pro Gly Arg 100 105 110 1105 110 1105 110 1105 110 110	Trp	Gly	Leu	Gln	Gly			Ser	Leu	Leu	Gly	Ala	Val	Tyr	Phe	80 Cys	
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Leu Gly Tyr Leu Leu Leu Gly Leu Leu Ala Met Leu Leu Ala Val Glu 130 131 135 Thr Phe Ser Glu Leu Pro Gln Val Arg Ala Met Gly Lys Phe Phe Arg 145 150 Pro Ser Gly Pro Val Thr Ala Glu Asp Gln Gly Gly Ile Leu Gly Gln 165 170 Asp Glu Leu Ala Leu Ser Thr Leu Pro Pro Ala Ala Pro Ala Ser Gly 180 180 181 Gln Ala Pro Ala Cys 195 <a blue"="" color:="" href="mailto:specific style=">specific style="color: blue">specific style="color: blue">sp	Gly	Arg	Ser	Leu	His	Pro	Val	Ile	Tyr	His	Leu	Gly	Gln	110 Leu	Ala	Leu	
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ceeegeege egegggeege etetegeega egetgtteee teegetteea ggtgtagege Met Thr Gly Gln Ser Leu Trp 1 5 gae gtg teg gag get aac gtc gag gac ggg gag atc ege atc aat gtg Asp Val Ser Glu Ala Asn Val Glu Asp Gly Glu Ile Arg Ile Asn Val 10 15 20 ggc ggc ttc aag agg agg etg ege teg eac acg etg etg ege ttc ecc Gly Gly Phe Lys Arg Arg Leu Arg Ser His Thr Leu Leu Arg Phe Pro 25 30 35 gag acg ege etg ggc ege ttg etg eac teg ege gag gec att 40 45 50 55 etg gag etc teg gat gac tac gac gac gtc eag egg gag ttc tac ttc Leu Glu Leu Cys Asp Asp Tyr Asp Asp Val Gln Arg Glu Phe Tyr Phe 60 65 70	ttca	gcac	cc a	agac	ccac	c ac	ggga	recto	gaco	CCCC	aaa	ctct	tgcc	ecc a	agcc	cagece	120
Met Thr Gly Gln Ser Leu Trp 1 Sac gtg tcg gag gct aac gtc gag gac ggg gag atc cgc atc aat gtg Asp Val Ser Glu Ala Asn Val Glu Asp Gly Glu Ile Arg Ile Asn Val 10 15 10 10 10 15 10 15 10 10		gcca	99 9	legea	cggc	gct	crcg	rccga	e cgc	tqtt	CCC	tcca	rctto	ca c	atai	agege	180 240
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gac gtg tcg gag gct aac gtc gag gac ggg gag atc cgc atc aat gtg Asp Val Ser Glu Ala Asn Val Glu Asp Gly Glu Ile Arg Ile Asn Val 10 15 20 ggc ggc ttc aag agg agg ctg cgc tcg cac acg ctg ctg cgc ttc ccc Gly Gly Phe Lys Arg Arg Leu Arg Ser His Thr Leu Leu Arg Phe Pro 25 30 35 gag acg cgc ctg ggc cgc ttg ctg ctc tgc cac tcg cgc gag gcc att Glu Thr Arg Leu Gly Arg Leu Leu Leu Cys His Ser Arg Glu Ala Ile 40 45 50 55 ctg gag ctc tgc gat gac tac gac gac gtc cag cgg gag ttc tac ttc Leu Glu Leu Cys Asp Asp Tyr Asp Asp Val Gln Arg Glu Phe Tyr Phe 60 65 70						:	_				rnr	GIY	, Gin		Let	ı Trp	
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ggc ggc ttc aag agg agg ctg cgc tcg cac acg ctg ctg cgc ttc ccc Gly Gly Phe Lys Arg Arg Leu Arg Ser His Thr Leu Leu Arg Phe Pro 25 30 35 gag acg cgc ctg ggc cgc ttg ctg ctc tgc cac tcg cgc gag gcc att 4 Glu Thr Arg Leu Gly Arg Leu Leu Leu Cys His Ser Arg Glu Ala Ile 40 45 50 55 ctg gag ctc tgc gat gac tac gac gcg gag gtc cag cgg gag ttc tac ttc 4 Leu Glu Leu Cys Asp Asp Tyr Asp Asp Val Gln Arg Glu Phe Tyr Phe 60 65 65 66 67 66 67 66 67 66 67 66 67 66 67 67 68 68	Asp	Val	Ser	gag Glu	gct Ala	aac Asn	gtc Val	gag Glu	gac Asp	ggg Gly	gag Glu	atc Ile	cgc Arq	atc Ile	aat Asn	gtg Val	342
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and the Arg Leu Gly Arg Leu Leu Cys His Ser Arg Glu Ala Ile 50 55 Ctg gag ctc tgc gat gac tac gac gac gtc cag cgg gag ttc tac ttc Leu Glu Leu Cys Asp Asp Tyr Asp Asp Val Gln Arg Glu Phe Tyr Phe 60 65 70 Gac cgc aac cct gag ctc ttc ccc tac gtg ctg cat ttc tat cac acc			*			:						7			•	•	
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Leu Glu Leu Cys Asp Asp Tyr Asp Asp Val Gln Arg Glu Phe Tyr Phe 60 65 70 gac cgc aac cct gag ctc ttc ccc tac gtg cat ttc tat cac acc			00			45		÷ .			,50∙	•					
60 65 70 Gao ogo aac oot gag oto tto oco tac gtg otg car tto tat cac acc		٠.	_														
gac ege aac eet gag ete tte eec tac gtg etg ear tte tat eac acc	ctg (gag	ctd	tgc g	gat	gac	tac	gac	gac	gtc	caq	caa .	gag	ttc	tac	ttc	486
gac ege aac eet gag ete tte eee tae gtg etg eat tte tat eac acc 5: Asp Arg Asn Pro Glu Leu Phe Pro Tyr Val Leu His Phe Tyr His Thr	ctg (Leu (gag Glu	ctc Leu	tgc (Cys)	gat . Asp .	gac Asp	tac Tyr	gac Asp	gac Asp	gtc Val	cag (Gln)	cgg (gag i	ttc Phe	tac	ttc	486
Asp Arg Asn Pro Glu Leu Phe Pro Tyr Val Leu His Phe Tyr His Thr	ctg (Leu (gag (Glu)	ctc Leu	tgc (Cys)	Asp .	gac Asp	tac Tyr	gac Asp	gac Asp	Val (cag Gln	cgg (gag i	Phe	tac Tyr	ttc	486
	beu (JIU .		cys /	Asp 60	Asp	ryr .	qeA	Asp	Val (Gln :	Arg	Glu 1	Phe	tac Tyr 70	ttc Phe	
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Arg Gln Gln Ile Arg Lys Ala Leu Asp Glu His Lys Glu Phe Lys Ala
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Glu Leu Ile Leu Tyr Arg Lys Ser Gly Leu Pro Phe Trp Cys Leu Leu
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                                                                    105
gat gtg ata ccc ata aag aat gag aaa ggg gag gtg gct ctc ttc cta
                                                                                                                                626
Asp Val Ile Pro Ile Lys Asn Glu Lys Gly Glu Val Ala Leu Phe Leu
                                                                  120
                                                                                   The state of the s
gte tet cae aag gae ate age gaa ace aag aac ega ggg gge eec gae
                                                                                                                               674
Val Ser His Lys Asp Ile Ser Glu Thr Lys Asn Arg Gly Gly Pro Asp
                      130
                                    135
aga tgg aaa gag aca ggt ggt ggc cgg cgc cga tat ggc cgg gca cga
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Arg Trp Lys Glu Thr Gly Gly Gly Arg Arg Arg Tyr Gly Arg Ala Arg
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His Leu Ser Gly His	: Leu Gln Lys Gln 180	ccc aag ggc aag cac Pro Lys Gly Lys His 185	Lys Leu 190
Asn Lys Gly Val Phe	Gly Glu Lys Pro	aac ttg cct gag tac Asn Leu Pro Glu Tyr 200	Lys Val 205
gcc gcc atc cgg aag Ala Ala Ile Arg Lys 210	tcg ccc ttc atc Ser Pro Phe Ile 215	ctg ttg cac tgt ggg Leu Leu His Cys Gly	gca ctg 914 Ala Leu
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acc acc tgg ttc ctg Thr Thr Trp Phe Leu 305	ctg gat gtc atc Leu Asp Val Ile 310	gca gcg ctg ccc ttt Ala Ala Leu Pro Phe 315	gac ctg 1202 Asp Leu
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acg gtg cgc ctg ctg Thr Val Arg Leu Leu 335	Arg Leu Leu Arg	ctg ctt ccg cgg ctg c Leu Leu Pro Arg Leu 345	gac cgg 1298 Asp Arg 350
tac tcg cag tac agc Tyr Ser Gln Tyr Ser 355	Ala Val Val Leu	aca ctg ctc atg gcc g Thr Leu Leu Met Ala N 360	gtg ttc 1346 Val Phe 365
gcc ctg ctc gcg cac Ala Leu Leu Ala His	Trp Val Ala Cys 375	gtc tgg ttt tac att g Val Trp Phe Tyr Ile (380	ggc cag 1394 Gly Gln
cgg gag atc gag agc	agc gaa tcc gag Ser Glu Ser Glu	ctg cct gag att ggc t Leu Pro Glu Ile Gly 7 395	tgg ctg 1442 Trp Leu
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tct Ser					! .				. • •	•					1. :	2162

 $\zeta_{i}\zeta^{i}$

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GIÀ	720	aat Asn	Thr	Leu	Met	Ser 725	Thr	Leu	Glu	Glu	Lys .730	Glu	Thr	Asp	Gly	2450
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Pro	ctg Leu	ctg Leu	tcc Ser	cct Pro 755	ggc	tgc Cys	acc Thr	tcc Ser	tca Ser 760	tcc Ser	tca Ser	gct Ala	gcc Ala	aag Lys 765	ctg Leu	2546
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aag																

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945	950		955		960
Pro His Pro Arg	Pro Gly 965	Pro Pro Pro	Leu Met Ala 970	Pro Arg Pro	Trp
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Trp::Thr Ser Thr				Gly Asp Leu	Cys
Ser Glu Pro Ser	Thr Pro	Ala Ser Pro			Ala
Arg Thr Gly Pro	Ala Glu				Thr
1025 Gly Glu Pro Pro	1030 Pro Gly			Pro Trp Asp	104 Pro
His Ser Leu Glu	1045 Met Val	Leu Ile Gly	1050 Cys His Gly	105 Ser Gly Thr	5 Val
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detection and control of the control	ettcagt ct ctaggg ga cgagta gt tttcct ag ctgggc tc atg atg Met Met 10 acg tct	cttagttc ca atgetttc tt ttttetgg tg ttatatat ge ttgaagac ge act gac cca Thr Asp Pro	gatgggtt ctc ccccagat ctt tttggcct cct tatcatat gtc atcactgg agc gtc aca tta Val Thr Leu 15 ttg acg cgt	tatggta ggaa tgecetg tagt tgttgg gtgg tgtttt etce ag atg gat a Met Asp A 1 aat gta ggt Asn Val Gly	tacagg 120 aggttt 180 aaaaag 240 tcttga 300 at gga 357 sn Gly gga 405 Gly 20 tcc 453
detection and categories and categor	ettcagt ct ctaggg ga cgagta gt tttcct ag ctgggc tc atg atg Met Met 10 acg tct	cttagttc ca atgetttc tt ttttetgg tg ttatatat ge ttgaagac ge act gac cca Thr Asp Pro	gatgggtt ctc ccccagat ctt tttggcct cct tatcatat gtc atcactgg agc gtc aca tta Val Thr Leu 15 ttg acg cgt Leu Thr Arg	tatggta ggaa ggcctg tagt ggtttt ctcc ag atg gat a Met Asp A 1 aat gta ggt Asn Val Gly tac ccg gat	tacagg 120 aggttt 180 aaaaag 240 tettga 300 at gga 357 sn Gly gga 405 Gly 20 tec 453 Ser
detection and control of the control	ettcagt ct ctaggg ga cgagta gt tttcct ag ctgggc tc atg atg Met Met 10 acg tct	cttagttc ca atgetttc tt ttttetgg tg ttatatat ge ttgaagac ge act gac cca Thr Asp Pro	gatgggtt ctc ccccagat ctt tttggcct cct tatcatat gtc atcactgg agc gtc aca tta Val Thr Leu 15 ttg acg cgt	tatggta ggaa tgecetg tagt tgttgg gtgg tgtttt etce ag atg gat a Met Asp A 1 aat gta ggt Asn Val Gly	tacagg 120 aggttt 180 aaaaag 240 tettga 300 at gga 357 sn Gly gga 405 Gly 20 tec 453 Ser
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inr	ASP	Pro	Val		reu	Asn	Val	GIY		His	Leu	Tyr	Thr	Thr	Ser	
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at a		363													÷	
Tan	mba	aca mb-	ttg	acg	-egr	Cac	ccg	gat	ECC	atg	CEE	gga	gct	atg	ttt	507
Leu	IIII	inr	Leu	inr	Arg	Tyr	Pro		ser	Met	Leu	GIY		Met	Phe	
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999	999	yac.	ttc	Dec	The	31-	cga	gac	CCE.	caa	ggc	aat	tac	ttt.	att	555
GIY	GIY	45	Phe	PIC	1111	Ald		Asp	Pro	Gin	GIY		Tyr	Phe	Ile	
		43					50					. 55				
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						65					/•		,			
tica	gaa	tta	асс	tta	CCO.	++~	ant.	-++	200		(r + + -	~~+	~+~			
Ser	Glu	Leu	Thr	Len	Pro	T.e.u	Aen	Dhe	Lve	Glu	Dhe	yac Nen	Tou	Tou	cgg	651
75	014	٠ ٠		neu	80	neu	vab	FILE	пуъ	85	PILE	ASP	Leu	reu	90	
	. •										• . •				30	
aaa	gaa	gca	gat	ttt	tac	cag	átt	дад	ccc	tta	art	cad	tert	ctc	aat	699
Lys	Glu	Ala	Asp	Phe	Tyr	Gln	Ile	Glu	Pro	Leu	Ile	Gln	Cvs	Leu	Asn	0,5
•			•	95	_				100				-,-	105		
	·														•	
gat	cct	aag	cct	ttg	tat	ccc	atg	gat	act	ttt	gaa	gaa	att	ata	gag	- 747
Asp	Pro	Lys	Pro	Leu	Tyr	Pro	Met	Asp	Thr	Phe	Glu	Glu	Val	Val	Glu	
-		_	110		-			115					120			
٠.	. /						.7					. •	٠.	٠.		
ctg	tct	agt	act	cgg	aag	ctt	tct	aag	tac	tcc	aac	ccá	gtg	gct	gtc	795
Leu	Ser	Ser	Thr	Arg	Lys	Leu	Ser	Lys	Tyr	Ser	Asn	Pro	Val	Ala	Val	
	•	125					130	•				135	: '			
					٠.,					: '`				•	•	
atc	ața	acg	caa	cta	acc	atc	acc	act.	aag	gte	cat	tcc	tta	cta	gaa	843
Ile	Ile	Thr	Gln	Leu	Thr	Ile	Thr	Thr	Lys	Val	His	Ser	Leu	Leu	Glu	
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			aat.													891
Gly	Ile	Ser	Asn	Tyr	Phe	Thr	Lys	Trp			Kis'	Met	Met	Asp	Thr	
155		٠,			160		•		•	165			•,	• •	170	
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Glu	Val	Ser	Leu	Arg	Val	His	Leu		Glu	Tyr	Ile		-	Gln	Gly	
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rne	Inr	тте	Arg	ASN	Inr	Arg	val	nis	HIE	Met	ser	GIU	Arg	Ala	Asn	

SUBJECT SUBJECT

· ·	205	210	215	
gaa aac Glu Asn 220	aca gtg gag c Thr Val Glu H	ac aac tgg act is Asn Trp Thr 225	ttc tgt agg cta Phe Cys Arg Leu 230	gcc cgg aag 1083 Ala Arg Lys
aca gac Thr Asp 235	gac t gatctcc Asp	gac cctgccacag	gttcctggaa agac	teteca 1133
ctcctcca tacctttt gcttttct ggctaaaa aaaacatc aaggaaaa caactgtt ctgactag	ty tittgitccc tt ataaaaagaa ta ttacagtgct at acaagaatga at gtagggtgac ct gttccaatca tc tttctatgca aa atatttatat gc ctggtgcagt	aggeagtgag gac ttccccctga gta gtctgaaaat cat aaaatgattt ctg aagaataagc aga ctagtttcca aac tttaaaagta ctt tataaatcaa gga tgaattctga ata ataattccca agt	cacagtg tgagatal cagaagg aagtttt cagaagg agtttt cagaagg gcctgtt cataggta tataact dataaaat ggtcccta gtactca tgatgcct caataaa taagtagt attaagt actgcttt accaaat atcctgtag gtactgt ctaccaga	gtg ctttggcaga 1253 cag agtctccaga 1313 cac ccttaacaga 1373 cac tcaactagaa 1433 ctt gagaaaaatc 1493 cat tgtaatatta 1553 ctt acagttatga 1613 gcc atggaaatgt 1673 ggt agaaaactta 1793
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gat aat ( Asp Asn (	gga gac tgg gg Gly Asp Trp Gl 5	tat atg atg y Tyr Met Met 10	act gac cca gtc Thr Asp Pro Val	aca tta aat 347
gta ggt ( Val Gly (	ga cac ttg ta	r Thr Thr Ser	ctc acc aca ttg Leu Thr Thr Leu 30	acg cgt tac 395 Thr Arg Tyr
ccg gat ( Pro Asp (	cc atg ctt gg Ser Met Leu Gl	a gct atg ttt g y Ala Met Phe ( 40	ggg ggg gac ttc Gly Gly Asp Phe 45	ccc aca gct 443 Pro Thr Ala
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cga tat o Arg Tyr \	ytc ctc aac tt Val Leu Asn Ph 70	c tta aga act t e Leu Arg Thr S	cca gaa ttg acc Ser Glu Leu Thr 75	tta ccg ttg 539 Leu Pro Leu 80

•	Asp	Phe	Lys	Glu 85	Phe	Asp	Leu	Leu	Arg 90	Lys	Glu	gca Ala	Asp	Phe 95	Tyr	Gln	587
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	acc Thr	act Thr	aag Lys	gtc Val	cat His 150	tee Ser	tta Leu	cta Leu	Glu	ggc Gly 155	atc Ile	tca Ser	aat Asn	tat Tyr	ttt Phe 160	acc Thr	779
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	agag	Tact	.ca t	gacg	JCCEL	t ga	igaaa	laato	aaa	acat	cat	gtag	igard	ac (	ctagt	ttcca	1438
	ctts	:ddtā itta:	idd D	aagt	agta.	t ac	caat	atta	aag	gaaa	act	gtto	caat	ca t	cttaa	aagta atcaa	1498
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	agto	tact	gt	tacc	agaa	a aa	aaaa	acaa	aac	taat	aaa	aaat	gaaa	ita t	gaaa	aaaaa	1738
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and sharping and the state

CENTERVOT COM

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							. ·	let A 1	sp A	sn G	ly A	sp T 5	tb G	ly T	yr Met		
atg Met 10	Inr	gac	cca Pro	gtc Val	aca Thr 15	Leu	aat Asn	gta Val	ggt Gly	gga Gly 20	cac His	ttg Leu	tat Tyr	aca Thr	acg Thr 25	16	12
ser	Leu	Inr	Thr	Leu 30	Thr	Arg	Tyr	ccg Pro	Asp 35	Ser	Met	Leu	Gly	Ala 40	Met	21	.0
Pne	GIŸ	GIĀ	45	hue	Pro	Tḥr	Ala	cga Arg 50	Asp	Pro	Gln	Gly	Asn 55	Tyr	Pne	25	8
TTE	Asp	Arg 60	Asp	GIY	Pro	Leu	Phe 65		Tyr	Val	Leu	Asn 70	Phe	Leu	Arg	30	6
rnr	75	GIU	Leu	Thr	Leu	Pro 80	Leu	gat Asp	Phe	Lys	Glu 85	Phe	Asp	Leu	Leu	35	4
90	гÀз	Giu	Ala	Asp	Phe 95	Tyr	Gln	att Ile	Glu	Pro 100	Leu	Ile	Gln	Cys	Leu 105	40	2
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gag Glu	ctg Leu	tct Ser	agt Ser 125	act Thr	cgg	aag Lys	ctt Leu	ser 130	aag Lys	tac Tyr	tcc Ser	aac Asn	cca Pro 135	gtg Val	gct Ala	498	8
gtc Val	atc Ile	ata Ile 140	acg Thr	caa Gln	cta Leu	acc Thr	atc Ile 145	acc Thr	act Thr	aag Lys	gtc Val	cat His 150	tcc Ser	tta Leu	cta Leu	546	6
gaa Glu	ggc Gly 155	atc Ile	tca Ser	aat Asn	tat Tyr	ttt Phe 160	acc Thr	aag Lys	tgg Trp	aat Asn	aag Lys 165	cac His	atg Met	atg Met	gac Asp	594	4
acc Thr 170	aga Arg	gac Asp	tgc Cys	cag Gln	gtt Val 175	tcc Ser	ttt Phe	act Thr	ttt Phe	gga Gly 180	ccc Pro	tgt Cys	gat Asp	tat Tyr	cac His 185	642	2
cag Gln	gaa Glu	gtt Val	tct Ser	ctt Leu 190	agg Arg	gtc Val	cac His	ctg Leu	atg Met 195	gaa Glu	tac Tyr	att Ile	aca Thr	aaa Lys 200	caa	690	)
ggt Gly	ttc Phe	acg Thr	atc Ile 205	cgc Arg	aac Asn	acc Thr	cgg Arg	gtg Val 210	cat His	cac His	atg Met	agt Ser	gagʻ Glu 215	cgg Arg	gcc Ala	738	3

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                                                                                                                                                                                             786
    Asn Glu Asn Thr Val Glu His Asn Trp Thr Phe Cys Arg Leu Ala Arg
                                                                               225
                                                                                                                                    230
    aag aca gac gac t gateteegae eetgeeacag gtteetggaa agacteteea
                                                                                                                                                                                            839
Lys Thr Asp Asp
    ggaaatggaa gatactgatt tttttttta aatcacagtg tgagatattt tttttcttt
                                                                                                                                                                                            899
    aaatagttgt atttatttga aggcagtgag gaccagaagg aagttttgtg ctttggcaga
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                                                                                                                                                                                         1319
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                                                                                                                                                                                         1542
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<210> 26 <211> 3204 in the control of the

 $e_{i}^{\mathrm{test}}(e_{i,j}) = e_{i,j}(e_{i,j})^{-1}$ 

		212>					٠.									
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	. <	22Ö>													*. *	
		221>	CDS	· · .	• • •										٠.;	
	<	222>	(18	2)	. (13	49)			•							
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•				•		•	•									
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aat	aaca	acc	regg	grgr	99 9	acag.	agtg	c gt	gcgt	gtgg	tgt	gtcc	cca	aggg	caggaa	60
gat	gaga	gga (	JJ49	cago	ta a	cgag	ataa	c gg	aggg. ctca	aggg ctca	gaa	gggc	999	agga	gaaaaa cagcgc	120
ga	tg a	gg c	उप व	to a	CC C	to t	tc c	tora.	ac o	oc a	gc c	cc a	gcy ac a	9999	cagege ga aag	180
M	et A	rg A	rg V	al T	hr L	eu Pi	he L	eu A	sn G	ly s	er P	ro L	vs A	sn G	ly Lys	229
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									٠.	,						
gtg	gtt	gct	gta	tat	gga	act	tta	tct	gat	ttg	ctt	tat	gtg	gcc	agc	277
Val	Val	ATG	20	lyr	GIY	inr	Leu		Asp	Leu	Leu	Ser			Ser	
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nea	50	Asp	Asp	TIE	Ala		IIe	Arg	Asp	Asp		Val	Leu	Phe	Val	
	7.0		.*			. 55		. *			60		<i>*</i> * * * * * * * * * * * * * * * * * *	. *		
tgt	gaa	gga	gaġ	ccá	ttt	att	gat	cct	cag	aca	gat	tct	aag	CCT	cct	421
Cys	Glu	Gly	Glu	Pro	Phė	Ile	Asp	Pro	Gln	Thr	Asp	Ser	Lvs	Pro	Pro	701
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							•									
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GIU	GIY	Deu	neu	85	PILE	nis	Inr	Asp	90	Leu	Thr	Leu	Asn		Gly	
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Gly	Arg	Tyr	Phe	Thr	Thr	Thr	Arg	Ser	Thr	Leu	Val	Asn	Lys	Glu	Pro	
		٠.	100			2 ., .		105		:	1		110			
~~~	aa+	2+~	ج نج	-à-												
Asp	Ser	Met	Len	Ala	Hig	Mar	Dhe	aag	gac	aaa	ggt	gtc Val	tgg	gga	aat	565
		115		ALU		1-16-0	120	Lys	Asp	гуs	GIY	125	Trp	GIY	Asn	
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Lys	Gln	Asp	His	Arg	Gly	Ala	Phe	Leu	Ile	Asp	Arg	Ser	Pro	Glu	Tyr	
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	<i>-</i>															
Phe	Glu	Pro	Tle	LLG	Agn	Tare	LEG	cgt	cat	gga	cag	ctc Leu	att	gta	aat	661
145				Deu	150	TYL	Deu	My	птр	155	GIN	ren	TTE	vaı	160	
							• '			4.7.7					100	
gat	ggc	att	aat	tta	ttg	ggt	gtg	tta	gaa	gaa	gca	aga	ttt	ttt	ggt	709
Asp	Gly	Ile	Asn	Leu	Leu	Gly	Val	Leu	Glu	Glu	Āla	Arg	Phe	Phe	Gly	-
	. 7			165					170			٠		175		
até	dec	t da ³	<u>- 4</u> -			a					:					_ = =
Tle	Agn	Ser	Len	Tle	Glu	CaC Wie	CCA	gaa	grg	gca	ata	aag Lys	aat	tct.	caa	757
			180	***	J_ u		TIE!	185	A Q. T	WIE,	TTE	_	190	ser	GIN	
	,		7/5 7							•			100			
cca	ccq	gaq	gat	cat	tca	cca	ata	tcc	caa	aag	gaa	ttt	atc	CC2	EEE	805

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Thr Gly Ile Asn Leu Arg Val Ala Thr Leu Lys Asn Ala Lys Leu Lys
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Phe Phe Leu Leu Gly Thr Thr Ile Leu Lys Pro Phe Met Leu Ser Ile
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Gln Lys Ala Leu Leu His Tyr Asn Glu Glu Ala Val Gln Ile Asn Pro
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Leu Ile Lys Lys 145	Tyr Asp Gln	Met Ala Ile Ph 150	c cac tgt tta ttt tgg e His Cys Leu Phe Trp 155	902
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V1032 6 4

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                                                   Met Arg Arg
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Gly Ala Leu Leu Ala Gly Ala Leu Ala Ala Tyr Ala Ala Tyr Leu Val
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Leu Gly Ala Leu Leu Val Ala Arg Leu Glu Gly Pro His Glu Ala Arg
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ctc cga gcc gag ctg gag acg ctg cgg gcg cag ctg ctt cag cgc agc
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Leu Arg Ala Glu Leu Glu Thr Leu Arg Ala Gln Leu Leu Gln Arg Ser
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Pro Cys Val Ala Ala Pro Ala Leu Asp Ala Phe Val Glu Arg Val Leu
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Ala Ser Thr Leu Ile Thr Thr Val Gly Tyr Gly Tyr Thr Thr Pro Leu
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Thr Asp Ala Gly Lys Ala Phe Ser Ile Ala Phe Ala Leu Leu Gly Val
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ceg ace ace atg ctg ctg ace gec tea gec eag ege etg tea etg
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Pro Thr Thr Met Leu Leu Leu Thr Ala Ser Ala Gln Arg Leu Ser Leu
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gac Asp	ccc Pro 165	Arg	cgg Arg	gcg Ala	gcc Ala	tgc Cys 170	tgg Trp	cac His	ttg Leu	gtg Val	gcc Ala 175	ctg Leu	ttg Leu	ej aaa	gtc Val	646
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Gin	Pro	tac Tyr 230	cgg Arg	gcc Ala	ctc Leu	Tyr	aag Lys 235	gtg Val	ctg Leu	gtc Val	aca Thr	gtc Val 240	Tyr	ctc Leu	ttc Phe	838
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tcc Ser 260	Asp	ctc Leu	cac His	ggc	ctc Leu 265	acg Thr	gag Glu	ctc Leu	atc Ile	ctg Leu 270	ctg Leu	ccc Pro	cct Pro	ccg Pro	tgc_ Cys 275	934
cct Pro	gcc Ala	agt Ser	ttc Phe	aat Asn 280	gcg Ala	gat Asp	gag Glu	gac Asp	gat Asp 285	cgg Arg	gtg Val	gac Asp	atc Ile	ctg Leu 290	ggc Gly	982
ccc Pro	cag Glŋ	ccg Pro	gag Glu 295	tcg Ser	cac His	cag Gln	caa Gln	ctc Leu 300	tct Ser	gcc Ala	agc Ser	tcc Ser	cac His 305	acc Thr	gac Asp	1030
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gtg Val	cct Pro	ggc Gly 325	Leu	gga Gly	ctg Leu	agg Arg	ggt Gly 330	cca Pro	Gly	gac Asp	cag Gln	agc Ser 335	tgg Trp	ctg Leu	tac Tyr	1126
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cgt Arg	ctc Leu	Ser 355	ttt Phe	Val	tcc Ser	Gln	cat His 360	ctg Leu	gct Ala	Gly ggg	atg Met	tga *	agg Arg 365			1222
ctc Leu	cct Pro	gtc Val 370	ccc Pro	atg Met	tcc Ser	Arg :	gct Ala 375	cca Pro	ctg Leu	ggc Gly	acc Thr	aac Asn 380	ata Ile	acc Thr	ttg Leu	1270
ttc	tct	gtc	ctt	tct	ctca	tcct	ct t	taca	ctgt	g to	tctc	tggc	tct	ctgg	cat	1325

Phe Ser Val Leu Ser 385

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 Leu Ser Leu Leu Leu Thr His Val Pro Leu Ser Trp Leu Ser Met Arg

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 160

 Trp Gly Trp Asp Pro Arg Arg Ala Ala Cys Trp His Leu Val Ala Leu
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 170
 175

 Leu Gly Val Val Val Thr Val Cys Phe Leu Val Pro Ala Val Ile Phe
 185
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 Ala His Leu Glu Glu Ala Trp Ser Phe Leu Asp Ala Phe Tyr Phe Cys
 205

 Phe Ile Ser Leu Ser Thr Ile Gly Leu Gly Asp Tyr Val Pro Gly Glu

210 215 220
Ala Pro Gly Gln Pro Tyr Arg Ala Leu Tyr Lys Val Leu Val Thr Val

225 230 235 Leu Val Thr Val

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Þ	lrg	His	Val	Ser 260		Leu	His	Gly	Leu 265	Thr	Glu	Leu	Ile	Leu 270	Leu	Pro	
			275					Asn 280					285	Arg	Val	_	
		290	•				295	Ser		,		300					
3	105					310		Pro			315					320	
					325	٠.		Leu		330	•		•		335	_	
				340				His	345					350			
			355					His 360				-	365				
		370			Ser	Arg	Ala 375	Pro	Leu	Gly		Asn 380	Ile	Thr	Leu	Phe	
³	er 85	Val	Leu	ser		•	٠	:		· . · · ·	·. ·						
			210> 211>		à.		ره ج	: :	٠, .		-			٠			
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			4 0 0 -	92				•	100							,	
а	aat		400> tgc (gcago	ct c	ggag	gege	age	ccgt	ctc	tgaa	ıtaaç			g gca	
а	aat				gcago			cgcgc	e ago	ccgt	ctc	tgaa	ıtaaç	Me	et Al	eg gca La Ala	
		geet	tge (ccgt	:	:	r .> ''						-	Me	et Al	a Ala	
c	ct	geet	tgc (ttg	cegt	gat	cct	aaa	tct	gcc	gct	cag	aac	tcc	Me :	et Al	a Ala	
c	ct	geet	tgc (ttg Leu	cegt	gat	cct	aaa		gcc	gct	cag	aac	tcc	Me :	et Al	a Ala	
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P	ct	gac Asp 5 /	ttg Leu ttt	ctg Leu tcc	gat Asp acg	cct Pro	aaa Lys 10	tct Ser	gcc Ala gtg	gct Ala	cag Gln gct	aac Asn 15	tcc Ser	aaa Lys	ccg Pro	agg Arg	
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o P	ct ro :tc eu 20	gac Asp 5 / tcg Ser	ttg Leu ttt Phe	ctg Leu tcc Ser	gat Asp acg Thr	cct Pro aaa Lys 25	aaa Lys 10 ccc Pro	tct Ser aca Thr	gcc Ala gtg Val	gct Ala ctt Leu	cag Gln gct Ala 30	aac Asn 15 tcc Ser	tcc Ser cgg Arg	aaa Lys gtg Val	ccg Pro gag Glu	agg Arg agt ser 35	106
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o P	ect Pro tcueu 20 accusp	gac Asp 5 / tcg Ser acg Thr	ttgc of ttgc o	ctg Leu tcc Ser att Ile	gat Asp acg Thr aat Asn 40	cct Pro aaa Lys 25 gtt Val	aaa Lys 10 ccc Pro atg Met	tct Ser aca Thr aaa Lys	gcc Ala gtg Val tgg Trp	gct Ala ctt Leu aag Lys 45	cag Gln gct Ala 30 acg Thr	aac Asn 15 tcc Ser gtc Val	tcc Ser cgg Arg tcc Ser	aaa Lys gtg Val acg Thr	ccg Pro gag Glu ata Ile 50	agg Arg agt Ser 35 ttc Phe	10 <i>6</i>
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o P	ect Pro tcueu 20 accusp	gac Asp 5 / tcg Ser acg Thr	ttgc of ttgc o	ctg Leu tcc Ser att Ile	gat Asp acg Thr aat Asn 40	cct Pro aaa Lys 25 gtt Val	aaa Lys 10 ccc Pro atg Met	tct Ser aca Thr aaa Lys	gcc Ala gtg Val tgg Trp	gct Ala ctt Leu aag Lys 45	cag Gln gct Ala 30 acg Thr	aac Asn 15 tcc Ser gtc Val	tcc Ser cgg Arg tcc Ser	aaa Lys gtg Val acg Thr	ccg Pro gag Glu ata Ile 50 aaa Lys	agg Arg agt Ser 35 ttc Phe	106 154 202
C P C L C L C L	ct Pro tc eu 20 ac sp	gac Asp 5 / tcg Ser acg Thr gtg Val	ttgc of ttgc o	ctg Leu tcc Ser att Ile gtc Val 55	gat Asp acg Thr aat Asn 40 ctc Leu	cct Pro aaa Lys 25 gtt Val tat Tyr	aaa Lys 10 ccc Pro atg Met ctg Leu	tct Ser aca Thr aaa Lys atc Ile	gcc Ala gtg Val tgg Trp atc Ile 60	gct Ala ctt Leu aag Lys 45 gga Gly	cag Gln gct Ala 30 acg Thr	aac Asn 15 tcc Ser gtc Val acc Thr	tcc Ser cgg Arg tcc Ser gtg Val	aaa Lys gtg Val acg Thr ttc Phe 65	ccg Pro gag Glu ata Ile 50 aaa Lys	agg Arg agt Ser 35 ttc Phe gca Ala	106 154 202
C P C L C L C L	ct Pro tc eu 20 ac sp	gac Asp 5 / tcg Ser acg Thr gtg Val	ttgc of ttgc of ttt ttt ttt ttt ttt ttt ttt ttt ttt	ctg Leu tcc Ser att Ile gtc Val 55	gat Asp acg Thr aat Asn 40 ctc Leu	cct Pro aaa Lys 25 gtt Val tat Tyr	aaa Lys 10 ccc Pro atg Met ctg Leu	tct Ser aca Thr aaa Lys atc Ile tca Ser	gcc Ala gtg Val tgg Trp atc Ile 60	gct Ala ctt Leu aag Lys 45 gga Gly	cag Gln gct Ala 30 acg Thr	aac Asn 15 tcc Ser gtc Val acc Thr	tcc Ser cgg Arg tcc Ser gtg Val att	aaa Lys gtg Val acg Thr ttc Phe 65	ccg Pro gag Glu ata Ile 50 aaa Lys	agg Arg agt Ser 35 ttc Phe gca Ala	106 154 202 250
C P C L C L C L	ct Pro tc eu 20 ac sp	gac Asp 5 / tcg Ser acg Thr gtg Val	ttgc of ttgc o	ctg Leu tcc Ser att Ile gtc Val 55	gat Asp acg Thr aat Asn 40 ctc Leu	cct Pro aaa Lys 25 gtt Val tat Tyr	aaa Lys 10 ccc Pro atg Met ctg Leu	tct Ser aca Thr aaa Lys atc Ile	gcc Ala gtg Val tgg Trp atc Ile 60	gct Ala ctt Leu aag Lys 45 gga Gly	cag Gln gct Ala 30 acg Thr	aac Asn 15 tcc Ser gtc Val acc Thr	tcc Ser cgg Arg tcc Ser gtg Val	aaa Lys gtg Val acg Thr ttc Phe 65	ccg Pro gag Glu ata Ile 50 aaa Lys	agg Arg agt Ser 35 ttc Phe gca Ala	106 154 202 250
OP OL SA	ect Pro etc 20 pac sp eu eu	gac Asp 5 / tcg Ser acg Thr gtg Val gag Glu caa	ttgc of ttgc o	ctg Leu tcc Ser att Ile gtc Val 55 cct Pro	gat Asp acg Thr aat Asn 40 ctc Leu cat His	cct Pro aaa Lys 25 gtt Val tat Tyr gag Glu	aaa Lys 10 ccc Pro atg Met ctg Leu att Ile	tct Ser aca Thr aaa Lys atc Ile tca Ser 75	gcc Ala gtg Val tgg Trp atc Ile 60 cag Gln	gct Ala ctt Leu aag Lys 45 gga Gly agg Arg	cag Gln gct Ala 30 acg Thr gcc Ala acc	aac Asn 15 tcc Ser gtc Val acc Thr	tcc Ser cgg Arg tcc Ser gtg Val att Ile 80	aaa Lys gtg Val acg Thr ttc Phe 65 gtg Val	ccg Pro gag Glu ata Ile 50 aaa Lys	agg Arg agt Ser 35 ttc Phe gca Ala cag Gln	106 154 202 250
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OP OL tl al	ict iccieu 20 iac icg icu icg icu	gac Asp 5 / tcg Ser acg Thr gtg Glu caa Gln 85	ttgc of ttgc o	ctg Leu tcc Ser att Ile gtc Val 55 cct Pro	gat Asp acg Thr aat Asn 40 ctc Leu cat His	cct Pro aaa Lys 25 gtt Val tat Tyr gag Glu tcc Ser	aaa Lys 10 ccc Pro atg Met ctg Leu att Ile caa Gln 90	tct Ser aca Thr aaa Lys atc ile tca Ser 75 cat	gcc Ala gtg Val tgg Trp atc Ile 60 cag Gln tcc Ser	gct Ala ctt Leu aag Lys 45 gga Gly agg Arg	cag Gln gct Ala 30 acg Thr gcc Ala acc Thr	aac Asn 15 tcc Ser gtc Val acc Thr acc Thr	tcc Ser cgg Arg tcc Ser gtg Val att Ile 80 tcg Ser	aaa Lys gtg Val acg Thr ttc Phe 65 gtg Val	ccg Pro gag Glu ata Ile 50 aaa Lys atc Ile	agg Arg agt Ser 35 ttc Phe gca Ala cag Gln	106 154 202 250

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Ser Phe Phe Phe Ala	Gly Thr Val Ile 140	aca acc ata gga ttt Thr Thr Ile Gly Phe 145	Gly Asn
Ile Ser Pro Arg Thr 150	Glu Gly Gly Lys 155	ata ttc tgt atc atc Ile Phe Cys Ile Ile 160	Tyr Ala
Leu Leu Gly Ile Pro 165	Leu Phe Gly Phe 170	ctc ttg gct gga gtt Leu Leu Ala Gly Val 175	Gly Asp
Gln Leu Gly Thr Ile 180	Phe Gly Lys Gly 185	att gcc aaa gtg gaa Ile Ala Lys Val Glu 190	Asp Thr 195
Phe Ile Lys Trp Asn 200	Val Ser Gln Thr	aag att cgc atc atc Lys Ile Arg Ile Ile 205	Ser Thr 210
Ile Ile Phe Ile Leu 215	Phe Gly Cys Val 220	ctc ttt gtg gct ctg Leu Phe Val Ala Leu 225	Pro Ala
Ile Ile Phe Lys His 230	Ile Glu Gly Trp 235	agt gcc ctg gac gcc Ser Ala Leu Asp Ala 240	Ile Tyr
Phe Val Val Ile Thr 245	Leu Thr Thr Ile 250	gga ttt ggt gac tac Gly Phe Gly Asp Tyr 255	Val Ala
Gly Gly Ser Asp Ile 260	Glu Tyr Leu Asp 265	ttc tat aag cct gtc Phe Tyr Lys Pro Val 270	Val Trp 275
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ttc cag cgg gcc acc Phe Gln Arg Ala Thr 340	tcc atc aag cgg Ser Ile Lys Arg 345	aag ctc tcg gca gaa Lys Leu Ser Ala Glu 350	ctg gct 1114 Leu Ala 355
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Gly Asn His Asn Gln Glu Leu Thr Pro Cys Arg Arg Thr Leu Ser Val
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                                                                 1210
Asn His Leu Thr Ser Glu Arg Asp Val Leu Pro Pro Leu Leu Lys Thr
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gag agt atc tat ctg aat ggt ttg acg cca cac tgt gct ggt gaa gag
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Glu Ser Ile Tyr Leu Asn Gly Leu Thr Pro His Cys Ala Gly Glu Glu
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att gct gtg att gag aac atc aaa tag ccctctctt aaataacctt
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Ile Ala Val Ile Glu Asn Ile Lys *
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International application No. PCT/US99/03826

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IPC(6) :	SSIFICATION OF SUBJECT MATTER CO7H 21/04; C07K 14/705; C12N 15/09, 15/63; C1 636/23.1, 24.3; 435/7.2, 69.1, 320.1; 530/350	2Q 1/68	
	o International Patent Classification (IPC) or to both	national classification and IPC	
	DS SEARCHED		
	ocumentation searched (classification system followe	ed by classification symbols)	
		d by classification symbolis;	
U.S. :	636/23.1, 24.3; 435/7.2, 69.1, 320.1; 530/350		
Documentati	ion searched other than minimum documentation to the	e extent that such documents are included	in the fields searched
El-amie de	to have accounted during the international according	Charles and the Name of Assault	
	ata base consulted during the international search (no Extra Sheet.	ame of data base and, where practicable,	, scarch terms used)
C. DOCI	UMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where ap	propriate, of the relevant passages	Relevant to claim No.
X,P	PARTISETI, M. et al. Cloning and Human Inward Rectifying Potassiu Expressed in Small Intestine. FEBS Let 176, see entire document.	um Channel Predominantly	1-9
·		·	
Furthe	or documents are listed in the continuation of Box C	See patent family annex.	
* Spe	eial estageries of cited documents:	"T" Inter document published after the inter	
	useant defining the general state of the art which is not considered to a functional relationship relations	date and not in conflict with the appli the principle or theory underlying the	
	ier document published on or after the internsticual filing date	"X" document of particular relevance; the	
"L" dom	unest which may throw doubts on priority claim(s) or which is to establish the publication date of enother citation or other sial reason (as specified)	occasidered novel or cannot be consider when the document is taken alone "Y" document of particular relevance; the	•
•	unest referring to an oral disclosure, use, exhibition or other	considered to involve an inventive combined with one or more other such being obvious to, a person skilled in th	step when the document is document, such combination
	ument published prior to the international filing date but leter then priority date claimed	*&* document member of the same patent	femily
Date of the a	octual completion of the international search	Date of mailing of the international scale 0 7 JUL 199	
Commission Box PCT	ailing address of the ISA/US or of Patents and Trademarks , D.C. 20231	Authorized Officer NIRMAL S. BASI	For
Facaimile No	o. (703) 305-3230	Telephone No. (703) 308-0196	

International application No. PCT/US99/03826

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, MEDLINE, JAPIO, BIOSIS, SCISEARCH, WPIDS, GENEMBL, NGENSEQ 34, EST, A-GENESEQ 32, PIR 58, SWISS-PROT 35, SPTREMBL 16.
search terms: potassium channel, K+hnov

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

Group I, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:2, the nucleic acid having the sequence of SEQ ID NO:1, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:2 and K+Hnov protein of SEQ ID NO:2.

Group II, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:4, the nucleic acid having the sequence of SEQ ID NO:3, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:4 and K+Hnov protein of SEQ ID NO:4.

Group III, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:6, the nucleic acid having the sequence of SEQ ID NO:5, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:6 and K+Hnov protein of SEQ ID NO:6.

Group IV, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:8, the nucleic acid having the sequence of SEQ ID NO:7, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:8 and K+Hnov protein of SEQ ID NO:8.

Group V, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:10, the nucleic acid having the sequence of SEQ ID NO:9, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:10 and K+Hnov protein of SEQ ID NO:10.

Group VI, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:12, the nucleic acid having the sequence of SEQ ID NO:11, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:12 and K+Hnov protein of SEQ ID NO:12.

Group VII, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:14, the nucleic acid having the sequence of SEQ ID NO:13, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:14 and K+Hnov protein of SEQ ID NO:14.

Group VIII, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:16, the nucleic acid having the sequence of SEQ ID NO:15, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:16 and K+Hnov protein of SEQ ID NO:16.

Group IX, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:18, the nucleic acid having the sequence of SEQ ID NO:17, nucleic acids hybridizing to said nucleic acids, expression cannot comprising said nucleic acids, cell comprising said expression cannot method for producing K+Hnov protein of SEQ ID NO:18 and K+Hnov protein of SEQ ID NO:18.

Group X, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:20, the nucleic acid having the sequence of SEQ ID NO:19, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:20 and K+Hnov protein of SEQ ID NO:20.

Group XI, claim(s)1-9, draws to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:25, the nucleic acid having the sequence of SEQ ID NO:21-25, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:25 and K+Hnov protein of SEQ ID NO:25.

Group XII, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:27, the aucleic acid having the sequence of SEQ ID NO:26, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing

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K+Hnov protein of SEQ ID NO:27 and K+Hnov protein of SEQ ID NO:27.

Group XIII, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:30, the nucleic acid having the sequence of SEQ ID NO:28-29, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:30 and K+Hnov protein of SEQ ID NO:30.

Group XIV, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:81, the nucleic acid having the sequence of SEQ ID NO:80, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:81 and K+Hnov protein of SEQ ID NO:81.

Group XV, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:83, the nucleic acid having the sequence of SEQ ID NO:82, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:83 and K+Hnov protein of SEQ ID NO:83.

Group XVI, claim(s)10, drawn to monoclonal antibody that binds to K+Hnov.

Group XVII, claim(s)11-14, drawn to non-human transgenic animal model for K+Hnov.

The inventions listed as Groups I-XVII do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: Group I is directed to nucleic acid (SEQ ID NO:1) encoding the K+Hnov protein of SEQ ID NO:2, nucleic acids hybridizing to raid nucleic acid, expression cassette comprising said nucleic acid, ceil comprising said cassette, method of producing the K+Hnov of SEQ ID NO:2 and the protein of SEQ ID NO:2. The special technical feature is the disclosed nucleic acid of SEQ ID NO:1 encoding the K+Hnov protein of SEQ ID NO:2. The nucleic acids, proteins, antibody and transgenic eximal model of Groups II-XVII do not share the special technical feature of Group I wherein the products of said Groups are structurally and functionally different. As above in Table 1, pages 8-9, the H+Nov proteins of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 25, 27, 30, 81 and 83 are all structurally and functionally different, the nucleic acids exceeding said proteins having different chromosome positions.

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International application No. PCT/US99/03826

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)	
This interv	national report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
ı. 🗀	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2.	Claims Nos.:
ل_ا	because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II C	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
	national Searching Authority found multiple inventions in this international application, as follows:
	case See Extra Sheet.
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} .	
ı. 🗀	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 9, SEQ ID NO:1 and 2
Romark o	The additional search fees were accompanied by the applicant's protest.
	No protest accompanied the new next of additional search fees.

Form PCT/ISA/210 (continuation of first sheet(1))(July 1992)*